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#### (57) Abstract

Methods for isolating K+Hnov genes are provided. The K+Hnov nucleic acid compositions find use in identifying homologous or related proteins and the DNA sequences encoding such proteins; in producing compositions that modulate the expression or function of the protein; and in studying associated physiological pathways. In addition, modulation of the gene activity in vivo is used for prophylactic and therapeutic purposes, such as identification of cell type based on expression, and the like.

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#### **HUMAN POTASSIUM CHANNEL GENES**

#### INTRODUCTION

#### Background

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lon channels are multi-subunit, membrane bound proteins critical for maintenance of cellular homeostasis in nearly all cell types. Channels are involved in a myriad of processes including modulation of action potentials, regulation of cardiac myocyte excitability, heart rate, vascular tone, neuronal signaling, activation and proliferation of T-cells, and insulin secretion from pancreatic islet cells. In humans, ion channels comprise extended gene families with hundreds, or perhaps thousands, of both closely related and highly divergent family members. The majority of known channels regulate the passage of sodium (Na\*), chloride (Cl\*), calcium (Ca\*\*) and potassium (K\*) ions across the cellular membrane.

Given their importance in maintaining normal cellular physiology, it is not surprising that ion channels have been shown to play a role in heritable human disease. Indeed, ion channel defects are involved in predisposition to epilepsy, cardiac arrhythmia (long QT syndrome), hypertension (Bartter's syndrome), cystic fibrosis, (defects in the CFTR chloride channel), several skeletal muscle disorders (hyperkalemic periodic paralysis, paramyotonia congenita, episodic ataxia) and congenital neural deafness (Jervell-Lange-Nielson syndrome), amongst others.

The potassium channel gene family is believed to be the largest and most diverse ion channel family. K\* channels have critical roles in multiple cell types andpathways, and are the focus of significant investigation. Four human conditions, episodic ataxia with myokymia, long QT syndrome, epilepsy and Bartter's syndrome have been shown to be caused by defective K\* ion channels. As the K\* channel family is very diverse, and given that these proteins are critical components of virtually all cells, it is likely that abnormal K\* channels will be involved in the etiology of additional renal, cardiovascular and central nervous system disorders of interest to the medical and pharmaceutical community.

The K\* channel superfamily can be broadly classified into groups, based upon the number of transmembrane domain (TMD) segments in the mature

protein. The minK (IsK) gene contains a single TMD, and although not a channel by itself, minK associates with different K\* channel subunits, such as KvLQT1 and HERG to modify the activity of these channels. The inward rectifying K+ channels (GIRK, IRK, CIR, ROMK) contain 2 TMD domains and a highly conserved pore domain. Twik-1 is a member of the newly emerging 4TMD K\* channel subset. Twik-like channels (leak channels) are involved in maintaining the steady-state K\* potentials across membranes and therefore the resting potential of the cell near the equilibrium potential for potassium (Duprat *et al.* (1997) EMBO J 16(17):5464-5471). These proteins are particularly intriguing targets for therapeutic regulation. The 6TMD, or Shaker-like channels, presently comprise the largest subset of known K\* channels. The slopoke (slo) related channels, or Ca\*\* regulated channels apparently have either 10 TMD, or 6 TMD with 4 additional hydrophobic domains.

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Four transmembrane domain, tandem pore domain K+ channels (4T/2P channels) represent a new family of potassium selective ion channels involved in the control of background membrane conductances. In mammals, five channels fitting the 4T/2P architecture have been described: TWIK, TREK, TASK-1, TASK-2 and TRAAK. The 4T/2P channels all have distinct characteristics, but are all thought to be involved in maintaining the steady-state K\* potentials across membranes and therefore the resting potential of the cell near the equilibrium potential for potassium (Duprat et al. (1997) EMBO J 16(17):5464-5471). These proteins are particularly intriguing targets for therapeutic regulation. Within this group, TWIK-1, TREK-1 and TASK-1 and TASK-2 are widely distributed in many different tissues, while TRAAK is present exclusively in brain, spinal cord and retina. The 4T/2P channels have different physiologic properties; TREK-1 channels, are outwardly rectifying (Fink et al. (1996) EMBO J 15(24):6854-62), while TWIK-1 channels, are inwardly rectifying (Lesage et al. (1996) EMBO J 15(5):1004-11. TASK channels are regulated by changes in PH while TRAAK channels are stimulated by arachidonic acid (Reyes et al. (1998) JBC 273(47):30863-30869).

The degree of sequence homology between different K\* channel genes is substantial. At the amino acid level, there is about 40% similarity between

different human genes, with distinct regions having higher homology, specifically the pore domain. It has been estimated that the K+ channel gene family contains approximately 10²-10³ individual genes. Despite the large number of potential genes, an analysis of public sequence databases and the scientific literature demonstrates that only a small number, approximately 20-30, have been identified. This analysis suggests that many of these important genes remain to be identified.

Potassium channels are involved in multiple different processes and are important regulators of homeostasis in nearly all cell types. Their relevance to basic cellular physiology and role in many human diseases suggests that pharmacological agents could be designed to specific channel subtypes and these compounds then applied to a large market (Bulman, D.E. (1997) Hum Mol Genet 6:1679-1685; Ackerman, M.J. and Clapham D.E. (1997) NEJM 336:1575-1586, Curran, M.E. (1998) Current Opinion in Biotechnology 9:565-572). The variety of therapeutic agents that modulate K+ channel activity reflects the diversity of physiological roles and importance of K+ channels in cellular function. A difficulty encountered in therapeutic use of therapeutic agents that modify K+ channel activity is that the presently available compounds tend to be non-specific and elicit both positive and negative responses, thereby reducing clinical efficacy. To facilitate development of specific compounds it is desirable to have further characterize novel K+ channels for use in *in vitro* and *in vivo* assays.

#### Relevant Literature

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A large body of literature exists in the general area of potassium channels. A review of the literature may be found in the series of books, "The Ion Channel Factsbook", volumes 1-4, by Edward C. Conley and William J. Brammar, Academic Press. An overview is provided of: extracellular ligand-gated ion channels (ISBN: 0121844501), intracellular ligand-gated channels (ISBN: 012184451X), Inward rectifier and intercellular channels (ISBN: 0121844528), and voltage gated channels (ISBN: 0121844536). Hille, B. (1992) "Ionic Channels of Excitable Membranes", 2<sup>nd</sup> Ed. Sunderland MA:Sinauer Associates, also reviews potassium channels.

Jan and Jan (1997) Annu. Rev. Neurosci. 20:91-123 review cloned potassium channels from eukaryotes and prokaryotes. Ackerman and Clapham (1997) N. Engl. J. Med. 336:1575-1586 discuss the basic science of ion channels in connection with clinical disease. Bulman (1997) Hum. Mol. Genet. 6:1679-1685 describe some phenotypic variation in ion channel disorders.

Stephan et al. (1994) Neurology 44:1915-1920 describe a pedigree segregating a myotonia with muscular hypertrophy and hyperirritability as an autosomal dominant trait (rippling muscle disease, Ricker et al. (1989) Arch. Neurol. 46405-408). Electromyography demonstrated that mechanical stimulation provoked electrically silent contractions. The responsible gene was localized to the distal end of the long arm of chromosome 1, in a 12-cM region near D1S235.

Type II pseudohypoaldosteronism is the designation used for a syndrome of chronic mineralocorticoid-resistant hyperkalemia with hypertension. The primary abnormality in type II PHA is thought to be a specific defect of the renal secretory mechanism for potassium, which limits the kaliuretic response to, but not the sodium and chloride reabsorptive effect of, mineralocorticoid. By analysis of linkage in families with autosomal dominant transmission, Mansfield *et al.* (1997) Nature Genet. 16:202-205 demonstrated locus heterogeneity of the trait, with linkage of the PHA2 gene to 1q31-q42 and 17p11-q21.

Sequences of four transmembrane, two pore potassium channels have been previously described. Reyes et al. (1998) J Biol Chem 273(47):30863-30869 discloses a pH sensitive channel. As with the related TASK-1 and TRAAK channels, the outward rectification is lost at high external K+ concentration. The TRAAK channel is described by Fink et al. (1998) EMBO J 17(12):3297-308. A cardiac two-pore channel is described in Kim et al. (1998) Circ Res 82(4):513-8. An open rectifier potassium channel with two pore domains in tandem and having a postsynaptic density protein binding sequence at the C terminal was cloned by Leonoudakis et al. (1998) J Neurosci 18(3):868-77.

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The electrophysiological properties of Task channels are of interest, (Duprat et al. (1997) EMBO J 16:5464-71). TASK currents are K+-selective, instantaneous and non-inactivating. They show an outward rectification when external [K+] is low, which is not observed for high [K+]out, suggesting a lack of

intrinsic voltage sensitivity. The absence of activation and inactivation kinetics as well as voltage independence are characteristic of conductances referred to as leak or background conductances. TASK is very sensitive to variations of extracellular pH in a narrow physiological range, a property probably essential for its physiological function, and suggests that small pH variations may serve a communication role in the nervous system.

## SUMMARY OF THE INVENTION

Isolated nucleotide compositions and sequences are provided for K+Hnov genes. The K+Hnov nucleic acid compositions find use in identifying homologous or related genes; in producing compositions that modulate the expression or function of its encoded proteins; for gene therapy; mapping functional regions of the proteins; and in studying associated physiological pathways. In addition, modulation of the gene activity *in vivo* is used for prophylactic and therapeutic purposes, such as treatment of potassium channel defects, identification of cell type based on expression, and the like

## DESCRIPTION OF THE SPECIFIC EMBODIMENTS

Nucleic acid compositions encoding *K+Hnov* polypeptides are provided. They are used in identifying homologous or related genes; in producing compositions that modulate the expression or function of the encoded proteins; for gene therapy; mapping functional regions of the proteins; and in studying associated physiological pathways. The *K+Hnov* gene products are members of the potassium channel gene family, and have high degrees of homology to known potassium channels. The encoded polypeptides may be alpha subunits, which form the functional channel, or accessory subunits that act to modulate the channel activity.

# CHARACTERIZATION OF K+HNOV

The sequence data predict that the provided K+Hnov genes encode potassium channels. Table 1 summarizes the DNA sequences, corresponding SEQ ID NOs, chromosomal locations, and polymorphisms. The provided

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sequences may encode a predicted K\*channel, e.g. voltage gated, inward rectifier, etc.; or a modulatory subunit.

Electrophysiologic characterization of ion channels is an important part of understanding channel function. Full length ion channel cDNAs may be combined with proper vectors to form expression constructs of each individual channel. Functional analyses of expressed channels can be performed in heterologous systems, or by expression in mammalian cell lines. For expression analyses in heterologous systems such as Xenopus oocytes, synthetic mRNA is made through in vitro transcription of each channel construct. mRNA is then injected, singly or in combination with interacting channel subunit mRNAs, into prepared oocytes and the cells allowed to express the channel for several days. Oocytes expressing the channel of interest are then analyzed by whole cell voltage clamp and patch clamp techniques.

To determine the properties of each channel when expressed in mammalian cells expression vectors specific to this type of analyses may be constructed and the resultant construct used to transform the target cells (for example human embryonic kidney (HEK) cells). Both stable and transiently expressing lines may be studied using whole cell voltage clamp and patch clamp techniques. Data obtained from EP studies includes, but is not limited to: current profiles elicited by depolarization and hyperpolarization, current-voltage (I-V) relationships, voltage dependence of activation, biophysical kinetics of channel activation, deactivation, and inactivation, reversal potential, ion selectivity, gating properties and sensitivity to channel antagonists and agonists.

Heterologous or mammalian cell lines expressing the novel channels can be used to characterize small molecules and drugs which interact with the channel. The same experiments can be used to assay for novel compounds which interact with the expressed channels.

In many cases the functional ion channel formed by K+Hnov polypeptides will be heteromultimers. Heteromultimers are known to form between different voltage gated, outward rectifying potassium channel  $\alpha$  subunits, generally comprising four subunits, and frequently associated with auxiliary,  $\beta$  subunits. Typically such  $\alpha$  subunits share a six-transmembrane domain structure (S1-S6),

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with one highly positively charged domain (S4) and a pore region situated between S5 and S6. Examples of such subunits are K+Hnov4, K+Hnov9, and K+Hnov12. Channels are also formed by mutimerization of subunits of the two transmembrane and one pore architecture. It is predicted that two subunits of K+Hnov49 or K+Hnov59 will be required to form a functional channel.

Heteromultimers of greatest interest are those that form between subunits expressed in the same tissues, and are a combination of subunits from the same species. In addition, the formation of multimers between the subject polypeptides and subunits that form functional channels are of particular interest. The resulting channel may have decreased or increased conductance relative to a homomultimer, and may be altered in response to beta subunits or other modulatory molecules.

Known voltage gated K+ channel α subunits include Kv1.1-1.8 (Gutman et al. (1993) Sem. Neurosci. 5:101-106); Kv2.1-2.2; Kv3.1-3.4; Kv4.1-4.3; Kv5.1; Kv6.1; Kv7.1; Kv8.1; Kv9.1-9.2. The subunits capable of forming ion inducing channels include all of those in the Kv1 through Kv4; and Kv7 families. The Kv5.1, Kv6.1, Kv8.1 and Kv9.1-9.2 subunits may be electrically silent, but functional in modifying the properties in heteromultimers.

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**TABLE 1** 

Name	CDNA SEQ	Protein SEQ	Polymorphisms	Chromosome Position	Channel Type
K+Hnov1	SEQ ID NO:1	SEQ ID NO:2	Alternative poly(A) tail: 1236, 2395	2q37	ATP-sensitive inward rectifying
K+Hnov4	SEQ ID NO:3	SEQ ID NO:4	A312C	unknown	Voltage gated K+ channel
			T335C		
			A377G		
	:		T344C	:	
: :	2. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1.		A401G		
			CA410-411GG (Ala/Thr)		
K+Hnov6	SEQ ID NO:5	SEQ ID NO:6		2p23	Delayed rectifying K+ channel
K+Hnov9	SEQ ID NO:7	SEC ID NO:8	Alternative poly(A) tail: 2304	8q23	Voltage gated K+ channel
K+Hnov12	SEQ ID NO:9	SEQ ID NO: 10	C321T (Pro/Leu)	Xp21	Voltage gated K+ channel
			A375G (GIWGIY)		
			C407T (LewPhe)		
K+Hnov15	SEQ ID NO:11	SEQ ID NO:12	Alternative poly(A) tail: 1427	13q14	modulatory subunit
			A689G (Gly/Arg)	#	\$
K+Hnov27	SEQ ID NO:13	SEQ ID NO:14	T365A (Ile/Asn)	18q12	modulatory subunit
K+Hnov2	SEQ ID NO:15	SEQ ID NO:16	N/A	NIA	4 transmembrane domain, 2 pore domain K+ channel

K+Hnov 11	SEQ ID NO:17	SEQ ID NO:18	NA	N/A	Human ortholog of murine gene, 6
			,		transmembrane dominas, voltage
	11				gated, delayed rectifier K+ channel
K+Hnov 14	SEQ ID NO:19	SEQ ID NO:20	C3166T	12q14	6 transmembrane domain, voltage gated K+ channel
K+Hnov28	SEQ ID NO:21-24	SEQ ID NO:25	4 aiternative 5' splices	3q29	Modulatory subunit
K+Hnov42	SEQ ID NO:26	SEQ ID NO:27	G1162A, T1460A, T2496A	8q11	Homology to K+ channel protein of C. elegans
K+Hnov44	SEQ ID NO:28-29	SEG ID NO:30	N/A	22p13	beta-subunit.
K'Hnov49	SEQ ID NO:80	SEQ ID NO.81	(ATCT), repeats in the 3' UTR sequence, starting at position 2166	1941	4T/2P channel, linked to the disease loci for rippling muscle disease 1 (RMD1), and type II pseudohypoaldosteronism
K'Hnov59	SEQ ID NO:82	SEQ ID NO:83	WA .	chr19	4T/2P channel

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# K+HNOV NUCLEIC ACID COMPOSITIONS

As used herein, the term "K+Hnov" is generically used to refer to any one of the provided genetic sequences listed in Table 1. Where a specific K+Hnov sequence is intended, the numerical designation, e.g. K49 or K59, will be added. Nucleic acids encoding K+Hnov potassium channels may be cDNA or genomic DNA or a fragment thereof. The term "K+Hnov gene" shall be intended to mean the open reading frame encoding any of the provided K+Hnov polypeptides, introns, as well as adjacent 5' and 3' non-coding nucleotide sequences involved in the regulation of expression, up to about 20 kb beyond the coding region, but possibly further in either direction. The gene may be introduced into an appropriate vector for extrachromosomal maintenance or for integration into a host genome.

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The term "cDNA" as used herein is intended to include all nucleic acids that share the arrangement of sequence elements found in native mature mRNA species, where sequence elements are exons and 3' and 5' non-coding regions. Normally mRNA species have contiguous exons, with the intervening introns, when present, removed by nuclear RNA splicing, to create a continuous open reading frame encoding a K+Hnov protein.

A genomic sequence of interest comprises the nucleic acid present between the initiation codon and the stop codon, as defined in the listed sequences, including all of the introns that are normally present in a native chromosome. It may further include the 3' and 5' untranslated regions found in the mature mRNA. It may further include specific transcriptional and translational regulatory sequences, such as promoters, enhancers, etc., including about 1 kb, but possibly more, of flanking genomic DNA at either the 5' or 3' end of the transcribed region. The genomic DNA may be isolated as a fragment of 100 kbp or smaller; and substantially free of flanking chromosomal sequence. The genomic DNA flanking the coding region, either 3' or 5', or internal regulatory sequences as sometimes found in introns, contains sequences required for proper tissue and stage specific expression.

The sequence of the 5' flanking region may be utilized for promoter elements, including enhancer binding sites, that provide for developmental regulation in tissues where *K+Hnov* genes are expressed. The tissue specific expression is useful for determining the pattern of expression, and for providing promoters that mimic the native pattern of expression. Naturally occurring polymorphisms in the promoter regions are useful for determining natural variations in expression, particularly those that may be associated with disease.

Alternatively, mutations may be introduced into the promoter regions to determine the effect of altering expression in experimentally defined systems. Methods for the identification of specific DNA motifs involved in the binding of transcriptional factors are known in the art, e.g. sequence similarity to known binding motifs, gel retardation studies, etc. For examples, see Blackwell et al. (1995) Mol Med 1: 194-205; Mortlock et al. (1996) Genome Res. 5: 327-33; and Joulin and Richard-Foy (1995) Eur J Biochem 232: 620-626.

The regulatory sequences may be used to identify *cis* acting sequences required for transcriptional or translational regulation of *K+Hnov* expression, especially in different tissues or stages of development, and to identify *cis* acting sequences and *trans* acting factors that regulate or mediate *K+Hnov* expression. Such transcription or translational control regions may be operably linked to a *K+Hnov* gene in order to promote expression of wild type or altered *K+Hnov* or other profeins of interest in cultured cells, or in embryonic, fetal or adult tissues, and for gene therapy.

The nucleic acid compositions of the subject invention may encode all or a part of the subject polypeptides. Double or single stranded fragments may be obtained of the DNA sequence by chemically synthesizing oligonucleotides in accordance with conventional methods, by restriction enzyme digestion, by PCR amplification, etc. For the most part, DNA fragments will be of at least 15 nt, usually at least 18 nt or 25 nt, and may be at least about 50 nt. Such small DNA fragments are useful as primers for PCR, hybridization screening probes, etc. Larger DNA fragments, i.e. greater than 100 nt are useful for production of the encoded polypeptide. For use in amplification reactions, such as PCR, a pair of

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primers will be used. The exact composition of the primer sequences is not critical to the invention, but for most applications the primers will hybridize to the subject sequence under stringent conditions, as known in the art. It is preferable to choose a pair of primers that will generate an amplification product of at least about 50 nt, preferably at least about 100 nt. Algorithms for the selection of primer sequences are generally known, and are available in commercial software packages. Amplification primers hybridize to complementary strands of DNA, and will prime towards each other.

The K+Hnov genes are isolated and obtained in substantial purity, generally as other than an intact chromosome. Usually, the DNA will be obtained substantially free of other nucleic acid sequences that do not include a K+Hnov sequence or fragment thereof, generally being at least about 50%, usually at least about 90% pure and are typically "recombinant", i.e. flanked by one or more nucleotides with which it is not normally associated on a naturally occurring chromosome.

The DNA may also be used to identify expression of the gene in a biological specimen. The manner in which one probes cells for the presence of particular nucleotide sequences, as genomic DNA or RNA, is well established in the literature and does not require elaboration here. DNA or mRNA is isolated from a cell sample. The mRNA may be amplified by RT-PCR, using reverse transcriptase to form a complementary DNA strand, followed by polymerase chain reaction amplification using primers specific for the subject DNA sequences. Alternatively, the mRNA sample is separated by gel electrophoresis, transferred to a suitable support, e.g. nitrocellulôse, nylon, etc., and then probed with a 25 fragment of the subject DNA as a probe. Other techniques, such as oligonucleotide ligation assays, in situ hybridizations, and hybridization to DNA probes arrayed on a solid chip may also find use. Detection of mRNA hybridizing to the subject sequence is indicative of K+Hnov gene expression in the sample.

The sequence of a K+Hnov gene, including flanking promoter regions and 30 cooling regions, may be mutated in various ways known in the art to generate targeted changes in promoter strength, sequence of the encoded protein, etc.

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The DNA sequence or protein product of such a mutation will usually be substantially similar to the sequences provided herein, i.e. will differ by at least one nucleotide or amino acid, respectively, and may differ by at least two but not more than about ten nucleotides or amino acids. The sequence changes may be substitutions, insertions or deletions. Deletions may further include larger changes, such as deletions of a domain or exon. Other modifications of interest include epitope tagging, e.g. with the FLAG system, HA, etc. For studies of subcellular localization, fusion proteins with green fluorescent proteins (GFP) may be used.

Techniques for *in vitro* mutagenesis of cloned genes are known. Examples of protocols for site specific mutagenesis may be found in Gustin *et al.*, *Biotechniques* 14:22 (1993); Barany, *Gene* 37:111-23 (1985); Colicelli *et al.*, *Mol Gen Genet* 199:537-9 (1985); and Prentki *et al.*, *Gene* 29:303-13 (1984). Methods for site specific mutagenesis can be found in Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*, CSH Press 1989, pp. 15.3-15.108; Weiner *et al.*, *Gene* 126:35-41 (1993); Sayers *et al.*, *Biotechniques* 13:592-6 (1992); Jones and Winistorfer, *Biotechniques* 12:528-30 (1992); Barton *et al.*, *Nucleic Acids Res* 18:7349-55 (1990); Marotti and Tomich, *Gene Anal Tech* 6:67-70 (1989); and Zhu, *Anal Biochem* 177:120-4 (1989). Such mutated genes may be used to study structure-function relationships of *K+Hnov*, or to alter properties of the protein that affect its function or regulation.

Homologs and orthologs of K+Hnov genes are identified by any of a number of methods. A fragment of the provided cDNA may be used as a hybridization probe against a cDNA library from the target organism of interest, where low stringency conditions are used. The probe may be a large fragment, or one or more short degenerate primers. Nucleic acids having sequence similarity are detected by hybridization under low stringency conditions, for example, at 50°C and 6XSSC (0.9 M sodium chloride/0.09 M sodium citrate) and remain bound when subjected to washing at 55°C in 1XSSC (0.15 M sodium chloride/0.015 M sodium citrate). Sequence identity may be determined by hybridization under stringent conditions, for example, at 50°C or higher and

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0.1XSSC (15 mM sodium chloride/01.5 mM sodium citrate). Nucleic acids having a region of substantial identity to the provided K+Hnov sequences, e.g. allelic variants, genetically altered versions of the gene, etc., bind to the provided K+Hnov sequences under stringent hybridization conditions. By using probes, particularly labeled probes of DNA sequences, one can isolate homologous or related genes. The source of homologous genes may be any species, e.g. primate species, particularly human; rodents, such as rats and mice, canines, felines, bovines, ovinas, equines, yeast, nematodes, etc.

Between mammalian species, e.g. human and mouse, homologs have 10 substantial sequence similarity, i.e. at least 75% sequence identity between nucleotide sequences, in some cases 80-or 90% sequence identity, and may be as high as 95% sequence identity between closely related species. Sequence similarity is calculated based on a reference sequence, which may be a subset of a larger sequence, such as a conserved motif, coding region, flanking region, etc. A reference sequence will usually be at least about 18 nt long, more usually at least about 30 nt long, and may extend to the complete sequence that is being compared. Algorithms for sequence analysis are known in the art, such as BLAST, described in Altschul et al. (1990), J. Mol. Biol. 215:403-10. In general, variants of the invention have a sequence identity greater than at least about 65%, preferably at least about 75%, more preferably at least about 85%, and may be greater than at least about 90% or more as determined by the Smith-Waterman homology search algorithm as implemented in MPSRCH program (Oxford Molecular). Exemplary search parameters for use with the MPSRCH program in order to identify sequences of a desired sequence identity are as follows: gap open penalty: 12; and gap extension penalty: 1. 25

## K+HNOV POLYPEPTIDES

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The subject nucleic acid sequences may be employed for producing all or portions of K+Hnov polypeptides. For expression, an expression cassette may be employed. The expression vector will provide a transcriptional and translational initiation region, which may be inducible or constitutive, where the coding region

is operably linked under the transcriptional control of the transcriptional initiation region, and a transcriptional and translational termination region. These control regions may be native to a K+Hnov gene; or may be derived from exogenous sources.

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The peptide may be expressed in prokaryotes or eukaryotes in accordance with conventional ways, depending upon the purpose for expression. For large scale production of the protein, a unicellular organism, such as *E. coli, B. subtilis, S. cerevisiae*, insect cells in combination with baculovirus vectors, or cells of a higher organism such as vertebrates, particularly mammals, *e.g.* COS 7 cells, may be used as the expression host cells. In some situations, it is desirable to express the *K+Hnov* gene in eukaryotic cells, where the *K+Hnov* protein will benefit from native folding and post-translational modifications. Small peptides can also be synthesized in the laboratory. Peptides that are subsets of the complete *K+Hnov* sequence may be used to identify and investigate parts of the protein important for function, or to raise antibodies directed against these regions.

Fragments of interest include the transmembrane and pore domains, the signal sequences, regions of interaction between subunits; etc. Such domains will usually include at least about 20 amino acids of the provided sequence, more usually at least about 50 amino acids, and may include 100 amino acids or more, up to the complete domain. Binding contacts may be comprised of non-contiguous sequences, which are brought into proximity by the tertiary structure of the protein. The sequence of such fragments may be modified through manipulation of the coding sequence, as described above. Truncations may be performed at the carboxy or amino terminus of the fragment, e.g. to determine the minimum sequence required for biological activity.

With the availability of the protein or fragments thereof in large amounts, by employing an expression host, the protein may be isolated and purified in accordance with conventional ways. A lysate may be prepared of the expression host and the lysate purified using HPLC, exclusion chromatography, get electrophoresis, affinity chromatography, or other purification technique. The

purified protein will generally be at least about 80% pure, preferably at least about 90% pure, and may be up to and including 100% pure. Pure is intended to mean free of other proteins, as well as cellular debris.

The expressed K+Hnov polypeptides are useful for the production of antibodies, where short fragments provide for antibodies specific for the particular polypeptide, and larger fragments or the entire protein allow for the production of antibodies over the surface of the polypeptide. Antibodies may be raised to the wild-type or variant forms of K+Hnov. Antibodies may be raised to isolated peptides corresponding to specific domains, e.g. the pore domain and the transmembrane domain, or to the native protein.

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Antibodies are prepared in accordance with conventional ways, where the expressed polypeptide or protein is used as an immunogen, by itself or conjugated to known immunogenic carriers, e.g. KLH, pre-S HBsAg, other viral or eukaryotic proteins, or the like. Various adjuvants may be employed, with a series of injections, as appropriate. For monoclonal antibodies, after one or more booster injections, the spleen is isolated, the lymphocytes immortalized by cell fusion, and then screened for high affinity antibody binding. The immortalized cells, i.e. hybridomas, producing the desired antibodies may then be expanded. For further description, see Monoclonal Antibodies: A Laboratory Manual, Harlow and Lane eds., Cold Spring Harbor Laboratories, Cold Spring Harbor, New York, 1988. If desired, the mRNA encoding the heavy and light chains may be isolated and mutagenized by cloning in E. coli, and the heavy and light chains mixed to further enhance the affinity of the antibody. Alternatives to in vivo immunization as a method of raising antibodies include binding to phage "display" libraries, usually in conjunction with in vitro affinity maturation.

#### K+HNOV GENOTYPING

The subject nucleic acid and/or polypeptide compositions may be used to genotyping and other analysis for the presence of polymorphisms in the sequence, or variation in the expression of the subject genes. Genotyping may be performed to determine whether a particular polymorphisms is associated with

a disease state or genetic predisposition to a disease state, particularly diseases associated with defects in excitatory properties of cells, e.g. cardiac, muscle, renal and neural cells. Disease of interest include rippling muscle disease, and type II psuedohypoaldosteronism.

Clinical disorders associated with K+ channel defects include long-QT syndrome; a congenital disorder affecting 1 in 10,000-15,000. Affected individuals have a prolonged QT interval in the electrocardiogram due to a delayed repolarization of the ventricle. Genetic linkage analyses identified two loci for long QT syndrome, LQT1, in 11p15.5 and LQT2, in 7q35-36. Positional cloning techniques identified the novel K+ channel KvLQT1 on chromosome 11 while candidate gene analysis identified causative mutations in the HERG K+ channel for LQT2.

The weaver mouse exhibits several abnormal neurological symptoms, including severe ataxia, loss of granule cell neurons in the cerebellum and dopaminergic cells in the substantia nigra, as well as seizures and male infertility. A G-protein-coupled K+ channel having a mutation in the conserved pore domain has been determined to cause the disease. The pancreatic-islet \(\mathcal{B}\)-cell ATP-sensitive K+ channel (KATP) is composed of two subunits, the sulfonylurea receptor (SUR) and the inward rectifier K+ channel Kir\(\mathcal{B}\)-2. Mutations in both SUR and Kir\(\mathcal{B}\)-2 have been identified in patients with persistent hyperinsulinemic hypoglycemia of infancy, which is caused by unregulated secretion of insulin.

Genotyping may also be performed for pharmacogenetic analysis to assess the association between an individual's genotype and that individual's ability to react to a therapeutic agent. Differences in target sensitivity can lead to toxicity or therapeutic failure. Relationships between polymorphisms in channel expression or specificity can be used to optimize therapeutic dose administration.

Genetic polymorphisms are identified in the K+Hnov gene (examples are listed in table 1), e.g. the repeat variation in the 3' UTR of K49. Nucleic acids comprising the polymorphic sequences are used to screen patients for altered reactivity and adverse side effects in response to drugs that act on K+ channels.

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K+Hnov genotyping is performed by DNA or RNA sequence and/or hybridization analysis of any convenient sample from a patient, e.g. biopsy material, blood sample, scrapings from cheek, etc. A nucleic acid sample from an individual is analyzed for the presence of polymorphisms in K+Hnov, particularly those that affect the activity, responsiveness or expression of K+Hnov. Specific sequences of interest include any polymorphism that leads to changes in basal expression in one or more tissues, to changes in the modulation of K+Hnov expression, or alterations in K+Hnov specificity and/or activity.

The effect of a polymorphism in K+Hnov gene sequence on the response to a particular agent may be determined by in vitro or in vivo assays. Such assays may include monitoring during clinical trials, testing on genetically defined cell lines, etc. The response of an individual to the agent can then be predicted by determining the K+Hnov genotype with respect to the polymorphism. Where there is a differential distribution of a polymorphism by racial background, 15 guidelines for drug administration can be generally tailored to a particular ethnic group.

Biochemical studies may be performed to determine whether a sequence polymorphism in a K+Hnov coding region or control regions is associated with disease, for example the association of K+Hnov 9 with idiopathic generalized epilepsy. Disease associated polymorphisms may include deletion or truncation of the gene, mutations that alter expression level, that affect the electrical activity of the channel, etc.

A number of methods are available for analyzing nucleic acids for the presence of a specific sequence. Where large amounts of DNA are available, genomic DNA is used directly. Alternatively, the region of interest is cloned into a suitable vector and grown in sufficient quantity for analysis. The nucleic acid may be amplified by conventional techniques, such as the polymerase chain reaction (PCR), to provide sufficient amounts for analysis. The use of the polymerase chain reaction is described in Saiki et al. (1985) Science 239:487, and a review of current techniques may be found in Sambrook et al. Molecular Cloning: A Laboratory Manual, CSH Press 1989, pp.14.2-14.33. Amplification may be used

to determine whether a polymorphism is present, by using a primer that is specific for the polymorphism. Alternatively, various methods are known in the art that utilize oligonucleotide ligation as a means of detecting polymorphisms, for examples see Riley et al. (1990) N.A.R. 18:2887-2890; and Delahunty et al. (1996) Am. J. Hum. Genet.58:1239-1246.

A detectable label may be included in an amplification reaction. Suitable labels include fluorochromes, e.g. fluorescein isothiocyanate (FITC), rhodamine, Texas Red, phycoerythrin, allophycocyanin, 6-carboxyfluorescein (6-FAM), 2',7'-dimethoxy-4',5'- dichloro-6-carboxyfluorescein (JOE), 6-carboxy-X-rhodamine (ROX), 6-carboxy-2',4',7',4,7- hexachlorofluorescein (HEX), 5-carboxyfluorescein (5-FAM) or N,N,N',N'-tetramethyl-6- carboxyrhodamine (TAMRA), radioactive labels, e.g. 32P, 35S, 3H; etc., The label may be a two stage system, where the amplified DNA is conjugated to biotin, haptens, etc. having a high affinity binding partner, e.g. avidin, specific antibodies, etc., where the binding partner is conjugated to a detectable label. The label may be conjugated to one or both of the primers. Alternatively, the pool of nucleotides used in the amplification is labeled, so as to incorporate the label into the amplification product.

The sample nucleic acid, e.g. amplified or cloned fragment, is analyzed by one of a number of methods known in the art. The nucleic acid may be sequenced by dideoxy or other methods. Hybridization with the variant sequence may also be used to determine its presence, by Southern blots, dot blots, etc. The hybridization pattern of a control and variant sequence to an array of oligonucleotide probes immobilised on a solid support, as described in U.S. 5,445,934, or in WO95/35505, may also be used as a means of detecting the presence of variant sequences. Single strand conformational polymorphism (SSCP) analysis, denaturing gradient gel electrophoresis (DGGE), mismatch cleavage detection, and heteroduplex analysis in gel matrices are used to detect conformational changes created by DNA sequence variation as alterations in electrophoretic mobility. Alternatively, where a polymorphism creates or destroys a recognition site for a restriction endonuclease (restriction fragment length polymorphism, RFLP), the sample is digested with that endonuclease, and the

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products size fractionated to determine whether the fragment was digested. Fractionation is performed by gel or capillary electrophoresis, particularly acrylamide or agarose gels.

In one embodiment of the invention, an array of oligonucleotides are provided, where discrete positions on the array are complementary to one or more of the provided sequences, e.g. oligonucleotides of at least 12 nt, frequently 20 nt, or larger, and including the sequence flanking a polymorphic position in a K\*Hnov sequence; coding sequences for different K\*Hnov channels, panels of ion channels comprising one or more of the provided K\* channels; etc. Such an array may comprise a series of oligonucleotides, each of which can specifically hybridize to a different polymorphism. For examples of arrays, see Hacia et al. (1996) Nature Genetics 14:441-447; Lockhart et al. (1996) Nature Biotechnol. 14:1675-1680; and De Risi et al. (1996) Nature Genetics 14:457-460.

Screening for polymorphisms in K+Hnov may be based on the functional or antigenic characteristics of the protein. Protein truncation assays are useful in detecting deletions that may affect the biological activity of the protein. Various immunoassays designed to detect polymorphisms in K+Hnov proteins may be used in screening. Where many diverse genetic mutations lead to a particular disease phenotype, functional protein assays have proven to be effective screening tools. The activity of the encoded K+Hnov protein as a potassium channel may be determined by comparison with the wild-type protein.

Antibodies specific for a K+Hnov may be used in staining or in immunoassays. Samples, as used herein, include biological fluids such as semen, blood, cerebrospinal fluid, tears, saliva, lymph, dialysis fluid and the like; organ or tissue culture derived fluids; and fluids extracted from physiological tissues. Also included in the term are derivatives and fractions of such fluids. The cells may be dissociated, in the case of solid tissues, or tissue sections may be analyzed. Alternatively a lysate of the cells may be prepared.

Diagnosis may be performed by a number of methods to determine the absence or presence or altered amounts of normal or abnormal K+Hnov polypeptides in patient cells. For example, detection may utilize staining of cells

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or histological sections, performed in accordance with conventional methods. The antibodies of interest are added to the cell sample, and incubated for a period of time sufficient to allow binding to the epitope, usually at least about 10 minutes. The antibody may be labeled with radioisotopes, enzymes, fluorescers, chemiluminescers, or other labels for direct detection. Alternatively, a second stage antibody or reagent is used to amplify the signal. Such reagents are well known in the art. For example, the primary antibody may be conjugated to biotin, with horseradish peroxidase-conjugated avidin added as a second stage reagent. Alternatively, the secondary antibody conjugated to a flourescent compound, e.g. flourescein, rhodamine, Texas red, etc. Final detection uses a substrate that undergoes a color change in the presence of the peroxidase. The absence or presence of antibody binding may be determined by various methods, including flow cytometry of dissociated cells, microscopy, radiography, scintillation counting, etc.

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## MODULATION OF GENE EXPRESSION

The K+Hnov genes, gene fragments, or the encoded protein or protein fragments are useful in gene therapy to treat disorders associated with K+Hnov defects. Expression vectors may be used to introduce the K+Hnov gene into a cell. Such vectors generally have convenient restriction sites located near the promoter sequence to provide for the insertion of nucleic acid sequences. Transcription cassettes may be prepared comprising a transcription initiation region, the target gene or fragment thereof, and a transcriptional termination region. The transcription cassettes may be introduced into a variety of vectors, e.g. plasmid; retrovirus, e.g. lentivirus; adenovirus; and the like, where the vectors are able to transiently or stably be maintained in the cells, usually for a period of at least about several days to several weeks.

The gene or K+Hnov protein may be introduced into tissues or host cells by any number of routes, including viral infection, microinjection, or fusion of vesicles. Jet injection may also be used for intramuscular administration, as 5

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described by Furth et al. (1992) Anal Biochem 205:365-368. The DNA may be coated onto gold microparticles, and delivered intradermally by a particle bombardment device, or "gene gun" as described in the literature (see, for example, Tang et al. (1992) Nature 356:152-154), where gold microprojectiles are coated with the K+Hnov or DNA, then bombarded into skin ceils.

Antisense molecules can be used to down-regulate expression of K+Hnov in cells. The anti-sense reagent may be antisense oligonuclectides (ODN), particularly synthetic ODN having chemical modifications from native nucleic acids, or nucleic acid constructs that express such anti-sense molecules as RNA. The antisense sequence is complementary to the mRNA of the targeted gene, and inhibits expression of the targeted gene products. Antisense molecules inhibit gene expression through various mechanisms, e.g. by reducing the amount of mRNA available for translation, through activation of RNAse H, or steric hindrance. One or a combination of antisense molecules may be administered, where a combination may comprise multiple different sequences.

Antisense molecules may be produced by expression of all or a part of the target gene sequence in an appropriate vector, where the transcriptional initiation is oriented such that an antisense strand is produced as an RNA molecule. Alternatively, the antisense molecule is a synthetic oligonucleotide. Antisense oligonucleotides will generally be at least about 7, usually at least about 12, more usually at least about 20 nucleotides in length, and not more than about 500, usually not more than about 50, more usually not more than about 35 nucleotides in length, where the length is governed by efficiency of inhibition, specificity, including absence of cross-reactivity, and the like. It has been found that short oligonucleotides, of from 7 to 8 bases in length, can be strong and selective inhibitors of gene expression (see Wagner et al. (1996) Nature Biotechnology 14:840-844).

A specific region or regions of the endogenous sense strand mRNA sequence is chosen to be complemented by the antisense sequence. Selection of a specific sequence for the oligonucleotide may use an empirical method, where several candidate sequences are assayed for inhibition of expression of

the target gene in an in vitro or animal model. A combination of sequences may also be used, where several regions of the rnRNA sequence are selected for antisense complementation.

Antisense oligonucleotides may be chemically synthesized by methods known in the art (see Wagner et al. (1993) supra. and Milligan et al., supra.) Preferred oligonucleotides are chemically modified from the native phosphodiester structure, in order to increase their intracellular stability and binding affinity. A number of such modifications have been described in the literature, which alter the chemistry of the backbone, sugars or heterocyclic 10 bases.

Among useful changes in the backbone chemistry are phosphorothioates; phosphorodithioates, where both of the non-bridging oxygens are substituted with sulfur; phosphoroamidites; alkyl phosphotriesters and boranophosphates. Achiral phosphate derivatives include 3'-O'-5'-S-phosphorothioate, 3'-S-5'-Ophosphorothicate, 3'-CH2-5'-O-phosphonate and 3'-NH-5'-O-phosphoroamidate. Peptide nucleic acids replace the entire ribose phosphodiester backbone with a peptide linkage. Sugar modifications are also used to enhance stability and affinity. The α-anomer of deoxyribose may be used, where the base is inverted with respect to the natural β-anomer. The 2'-OH of the ribose sugar may be altered to form 2'-O-methyl or 2'-O-allyl sugars, which provides resistance to degradation without comprising affinity. Modification of the heterocyclic bases must maintain proper base pairing. Some useful substitutions include deoxyuridine for deoxythymidine; 5-methyl-2'-deoxycytidine and 5-bromo-2'deoxycytidine for deoxycytidine. 5- propynyl-2'-deoxyuridine and 5-propynyl-2'-25 deoxycytidine have been shown to increase affinity and biological activity when substituted for deoxythymidine and deoxycytidine, respectively.

As an alternative to anti-sense inhibitors, catalytic nucleic acid compounds, e.g. ribozymes, anti-sense conjugates, etc. may be used to inhibit gene expression. Ribozymes may be synthesized in vitro and administered to the 20 patient, or may be encoded on an expression vector, from which the ribozyme is synthesized in the targeted cell (for example, see International patent application

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WO 9523225, and Beigelman et al. (1995) Nucl. Acids Res 23:4434-42). Examples of oligonucleotides with catalytic activity are described in WO 9506764. Conjugates of anti-sense ODN with a metal complex, e.g. terpyridylCu(II), capable of mediating mRNA hydrolysis are described in Bashkin et al. (1995) Appl 5 Biochem Biotechnol 54:43-56.

# GENETICALLY ALTERED CELL OR ANIMAL MODELS FOR K+HNOV FUNCTION

The subject nucleic acids can be used to generate transgenic animals or site specific gene modifications in cell lines. Transgenic animals may be made through homologous recombination, where the normal K+Hnov locus is altered. Alternatively, a nucleic acid construct is randomly integrated into the genome. Vectors for stable integration include plasmids, retroviruses and other animal viruses, YACs, and the like.

The modified cells or animals are useful in the study of K+Hnov function and regulation. For example, a series of small deletions and/or substitutions may be made in the K+Hnov gene to determine the role of different transmembrane domains in forming multimeric structures, ion channels, etc. Of interest are the use of K+Hnov to construct transgenic animal models for epilepsy and other neurological defects, where expression of K+Hnov is specifically reduced or absent. Specific constructs of interest include anti-sense K+Hnov, which will block K+Hnov expression, expression of dominant negative K+Hnov mutations, etc. One may also provide for expression of the K+Hnov gene or variants thereof in cells or tissues where it is not normally expressed or at abnormal times of development.

DNA constructs for homologous recombination will comprise at least a portion of the K+Hnov gene with the desired genetic modification, and will include regions of homology to the target locus. DNA constructs for random integration need not include regions of homology to mediate recombination. Conveniently, markers for positive and negative selection are included. Methods for generating cells having targeted gene modifications through homologous recombination are

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known in the art. For various techniques for transfecting mammalian cells, see Keown et al. (1990) Methods in Enzymology 185:527-537.

For embryonic stem (ES) cells, an ES cell line may be employed, or embryonic cells may be obtained freshly from a host, e.g. mouse, rat, guinea pig, etc. Such cells are grown on an appropriate fibroblast-feeder layer or grown in the presence of leukemia inhibiting factor (LIF). When ES or embryonic cells have been transformed, they may be used to produce transgenic animals. After transformation, the cells are plated onto a feeder layer in an appropriate medium. Cells containing the construct may be detected by employing a selective medium. After sufficient time for colonies to grow, they are picked and analyzed for the occurrence of homologous recombination or integration of the construct. Those colonies that are positive may then be used for embryo manipulation and blastocyst injection. Blastocysts are obtained from 4 to 6 week old superovulated females. The ES cells are trypsinized, and the modified cells are injected into the blastocoel of the blastocyst. After injection, the blastocysts are returned to each uterine horn of pseudopregnant females. Females are then allowed to go to term and the resulting offspring screened for the construct. By providing for a different phenotype of the blastocyst and the genetically modified cells, chimeric progeny can be readily detected.

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The chimeric animals are screened for the presence of the modified gene and males and females having the modification are mated to produce homozygous progeny. If the gene alterations cause lethality at some point in development, tissues or organs can be maintained as allogeneic or congenic grafts or transplants, or in *in vitro* culture. The transgenic animals may be any non-human mammal, such as laboratory animals, domestic animals, etc. The transgenic animals may be used in functional studies, drug screening, etc., e.g. to determine the effect of a candidate drug on Ras or related gene activation, oncogenesis, etc.

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# TESTING OF K+HNOV FUNCTION and RESPONSES

Potassium channels such as K+Hnov polypeptides are involved in multiple biologically important processes. Pharmacological agents designed to affect only specific channel subtypes are of particular interest. Presently available compounds tend to be non-specific and elicit both positive and negative responses, thereby reducing clinical efficacy.

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The subject polypeptides may be used in *in vitro* and *in vivo* models to test the specificity of novel compounds, and of analogs and derivatives of compounds known to act on potassium channels. Numerous pharmacological agents have profound affects on K+ channel activity. As examples, Sotalol (BETAPACE) is a class III antiarrhythmic drug that prolongs cardiac action potentials by inhibiting delayed rectifier K+ channels. Sulfonylurea drugs, such as Glipizide (GLUCOTROL) and Tolazamide (TOLAMIDE) function as antidiabetic drugs by blocking ATP-sensitive K+ channels present in pancreatic islet cells, thereby regulating insulin secretion. Diazoxide (HYPERSTAT IV) is an antihypertensive drug that activates ATP-sensitive K+ channels, resulting in the relaxation of vascular smooth muscle. There are several other examples of drugs that have antidiabetic, antihypertensive, or antiarrhythmic activities. A number of drugs that activate K+ channels that have been proposed as coronary vasodilators for the treatment of both vasospastic and chronic stable angina.

The availability of multiple K+ channel subunits allows *in vitro* reconstruction of functional channels, which may comprise different alpha and beta subunits. The individual components may be modified by sequence deletion, substitution, *etc.* to determine the functional role of specific domains.

Drug screening may be performed using an *in vitro* model, a genetically altered cell or animal, or purified K+Hnov protein, either as monomers, homomultimers or hetermultimers. One can identify ligands or substrates that bind to, modulate or mimic the action of K+Hnov. Drug screening identifies agents that provide a replacement for K+Hnov function in abnormal cells. Of particular interest are screening assays for agents that have a low toxicity for human cells. A wide variety of assays may be used for this purpose, including

monitoring cellular excitation and conductance, labeled *in vitro* protein-protein binding assays, electrophoretic mobility shift assays, immunoassays for protein binding, and the like. The purified protein may also be used for determination of three-dimensional crystal structure, which can be used for modeling intermolecular interactions.

The term "agent" as used herein describes any molecule, e.g. protein or pharmaceutical, with the capability of altering or mimicking the physiological function of K+Hnov polypeptide. Generally a plurality of assay mixtures are run in parallel with different agent concentrations to obtain a differential response to the various concentrations. Typically, one of these concentrations serves as a negative control, i.e. at zero concentration or below the level of detection.

Candidate agents encompass numerous chemical classes, though typically they are organic molecules, preferably small organic compounds having a molecular weight of more than 50 and less than about 2,500 daltons. Candidate agents comprise functional groups necessary for structural interaction with proteins, particularly hydrogen bonding, and typically include at least an amine, carbonyl, hydroxyl or carboxyl group, preferably at least two of the functional chemical groups. The candidate agents often comprise cyclical carbon or heterocyclic structures and/or aromatic or polyaromatic structures substituted with one or more of the above functional groups. Candidate agents are also found among biomolecules including peptides, saccharides, fatty acids, steroids, purines, pyrimidines, derivatives, structural analogs or combinations thereof.

Candidate agents are obtained from a wide variety of sources including libraries of synthetic or natural compounds. For example, numerous means are available for random and directed synthesis of a wide variety of organic compounds and biomolecules, including expression of randomized oligonucleotides and oligopeptides. Alternatively, libraries of natural compounds in the form of bacterial, fungal, plant and animal extracts are available or readily produced. Additionally, natural or synthetically produced libraries and compounds are readily modified through conventional chemical, physical and biochemical means, and may be used to produce combinatorial libraries. Known

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pharmacological agents may be subjected to directed or random chemical modifications, such as acylation, alkylation, esterification, amidification, etc. to produce structural analogs.

Where the screening assay is a binding assay, one or more of the molecules may be joined to a label, where the label can directly or indirectly provide a detectable signal. Various labels include radioisotopes, fluorescers, chemiluminescers, enzymes, specific binding molecules, particles, e.g. magnetic particles, and the like. Specific binding molecules include pairs, such as biotin and streptavidin, digoxin and antidigoxin etc. For the specific binding members, the complementary member would normally be labeled with a molecule that provides for detection, in accordance with known procedures.

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A variety of other reagents may be included in the screening assay. These include reagents like salts, neutral proteins, e.g. albumin, detergents, etc that are used to facilitate optimal protein-protein binding and/or reduce non-specific or background interactions. Reagents that improve the efficiency of the assay, such as protease inhibitors, nuclease inhibitors, anti-microbial agents, etc. may be used. The mixture of components are added in any order that provides for the requisite binding. Incubations are performed at any suitable temperature, typically between 4 and 40°C. Incubation periods are selected for optimum activity, but may also be optimized to facilitate rapid high-throughput screening. Typically between 0.1 and 1 hours will be sufficient.

The compounds having the desired pharmacological activity may be administered in a physiologically acceptable carrier to a host in a variety of ways, orally, topically, parenterally e.g. subcutaneously, intraperitoneally, by viral infection, intravascularly, etc. Depending upon the manner of introduction, the compounds may be formulated in a variety of ways. The concentration of therapeutically active compound in the formulation may vary from about 0.1-100 wt.%. The pharmaceutical compositions can be prepared in various forms, such as granules, tablets, pills, suppositories, capsules, suspensions, salves, lotions and the like. Pharmaceutical grade organic or inorganic carriers and/or diluents suitable for oral and topical use can be used to make up

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compositions containing the therapeutically-active compounds. Diluents known to the art include aqueous media, vegetable and animal oils and fats. Stabilizing agents, wetting and emulsifying agents, salts for varying the osmotic pressure or buffers for securing an adequate pH value, and skin penetration enhancers can be used as auxiliary agents.

It is to be understood that this invention is not limited to the particular methodology, protocols, cell lines, animal species or genera, and reagents described, as such may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims.

As used herein the singular forms "a", "and", and "the" include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to "a cell" includes a plurality of such cells and reference to "the cell" includes reference to one or more cells and equivalents thereof known to those skilled in the art, and so forth. All technical and scientific terms used herein have the same meaning as commonly understood to one of ordinary skill in the art to which this invention belongs unless clearly indicated otherwise.

It must be noted that as used herein and in the appended claims, the singular forms "a", "and", and "the" include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to "a complex" includes a plurality of such complexes and reference to "the formulation" includes reference to one or more formulations and equivalents thereof known to those skilled in the art, and so forth.

All publications mentioned herein are incorporated herein by reference for the purpose of describing and disclosing, for example, the methods and methodologies that are described in the publications which might be used in connection with the presently described invention. The publications discussed above and throughout the text are provided solely for their disclosure prior to the filing date of the present application. Nothing herein is to be construed as an

admission that the inventors are not entitled to antedate such disclosure by virtue of prior invention.

## EXPERIMENTAL

The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how to make and use the subject invention, and are not intended to limit the scope of what is regarded as the invention. Efforts have been made to ensure accuracy with respect to the numbers used (e.g. amounts, temperature, concentrations, etc.) but some experimental errors and deviations should be allowed for. Unless otherwise indicated, parts are parts by weight, molecular weight is average molecular weight, temperature is in degrees centigrade; and pressure is at or near atmospheric.

#### 15 Methods

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Two different types of sequence searches were performed. The first centered on the most highly conserved region of the K+ channel family, the pore domain. The pore is composed of 15-17 amino acids and can be divided into subfamilies based on the number of transmembrane segments present in the channel. Eleven variant peptide sequences corresponding to the pore domain were used in TBLASTN searches against the EST division of Genbank. Significant matches were identified, and classified into 2 categories: identical to known human K+ channels and related to known K+ channels. The pore sequences are shown in Table 2.

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# TABLE

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SEQ ID NO	Genbank #	
49	L02751	TGGTGGGCTGTGGTGACCATGACAACTGTGGGCTATGGGGACATG
50	M60451	TGGTGGGCAGTGGTCACCATGACCACTGTGGGCTACGGGGGACATG
51	L02752	TGGTGGGCAGTCGTCCATGACAACTGTAGGCTATGGAGACATG
52	M55515	TGGTGGGCAGTGGTAACCATGACAACAGTGGGTTACGGCGATATG
53	211585	TGGTGGGCTGTGGTCACCATGACGACCCTGGGCTATGGAGACATG
55	U40990	TGGTGGGGGTGGTCACAGTCACCATCGGCTATGGGGACAAG
55	126643	TGGTGGCCAGTGGCCATGACCACGGTTGGCTATGGGGACATG
56	M96747	TGGTGGGCCGTGGTCACCATGACGACCCTGGGCTATGGAGACATG
57	M64676	TGGTGGGCTGTGGTCACCATGACGACACTGGGCTACGGAGACATG
58	M55514	TGGTGGGCTGTGGTGACCATGTGGGGCTATGGGGACATG
59	X83582	TTCCTGTTCTCCATTGAGACCGAAACAACCATTGGGTATGGCTTCCG
99	578684	TTTTTATTCTCAATAGAGACAGAACCACCATTGGTTATGGCTACCG
19	U22413	TTCCTCTTCTCCATTGAGACCCAGACCATAGGCTATGGTTTCAG
62	U24056	TTCCTGTTCTCGGTGGAGGCGCAGACGACCATCGGCTATGGGTTCCG
63	U52155	TTCCTCTTCTCCCTTGAATCCCAAACCACCATTGGCTATGGCTTCCG
2	D87281	TITCICITITICCCTGGAATCCCAGACAACCATTGGCTATGGAGTCCG
	D50582	TTCCTTTTCTCCATTGAGGTCCAAGTGACTATTGGCTTTGGGGGGCG
88	D50315	THETETTETECATTGAAGTTCAAGTTACCATTGGGTTTGGAGGGAG
67	U04270	GCGCTCTACTTCACCTTCAGCAGCCTCACCAGTGTGGGCTTCGGCAC

The unique pore peptides sequences are shown in Table 3.

TABLE 3

	TABLE 3
SEQ ID NO	Amino acid sequence
68	WWAVVSMTTVGYGDM
69	WWAVVTMTTLGYGDM
70	WWGWTVTTIGYGDK
71	WWAVVTMTTVGYGDM
72	FLFSIEVQVTIGFGG
73	FLFSLESQTTIGYGV
74	FLFSIETETTIGYGY
75	FLFSIETQTTIGYGF
76	FLFSVETQTTIGYGF
77	FLFSLESQTTIGYGF
78	FLFSIETETTIGYGF
79	ALYFTFSSLTSVGFGN

The second set of experiments was based on a complex, reiterative process.

Annotated protein and DNA sequences were obtained from GenBank for all known K+ channels from all species. The TBLASTN and BLASTN programs were used to identify homologous ESTs, which were then analyzed using the BLASTX and BLASTN algorithms to identify ESTs which were related to K+ channels yet not identical to any known human K+ channel gene.

Novel human K+ channels were defined as those that had clear homology to known K+ channels from any species and were not present as identities or near identities to any human-derived sequences in any division of Genbank.

Isolation of full length cDNA sequence. EST clones were picked from the IMAGE consortium cDNA library and end-sequenced with vector primers. Gap closure was achieved either by primer walking or transposon sequencing. GeneTrapper (Life

Technologies) was used to isolate larger cDNA clones according to the provided protocol. RACE was used to extend the sequences as necessary using standard protocols.

Sequences were assembled in Sequencher (Gene Codes). The presence of open reading frames was assessed as well as potential start codons. Potential polymorphisms were detected as sequence variants between multiple independent clones. Sequence homologies were detected using the BLAST algorithms.

The completed gene sequences and predicted amino acid sequences are provided as SEQ ID NO:1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21-24, 26 and 28-29. Polymorphisms, chromosome locations and family assignments are shown in Table 1.

ESTs that had top human hits with >95% identity over 100 amino acids were discarded. This was based upon the inventors' experience that these sequences were usually identical to the starting probe sequences, with the differences due to sequence error. The remaining BLASTN and BLASTX outputs for each EST were examined manually, i.e., ESTs were removed from the analysis if the inventors determined that the variation from the known related probe sequence was a result of poor database sequence. Poor database sequence was usually identified as a number of 'N' nucleotides in the database sequence for a BLASTN search and as a base deletion or insertion in the database sequence, resulting in a peptide frameshift, for a BLASTX output. ESTs for which the highest scoring match was to non-related sequences were also discarded at this stage. The EST sequences that correspond to each clone are shown in Table 4.

Table 4 January 1997 St. A. A. A. A.

Genbank Accession#			Trace "	IMAGE Plate Coordinates	Read 5'/3'
N39619	K+Hnov2	277113	yy51h05.s1	611p10	3'
N46767	K+Hnov2	277113	yy51h05.r1	611p10	5'
R19352	K+Hnov11	33144	yg24f12.r1	155024	5'
R44628	K+Hnov11	33144	yg24f12.s1	155024	3'

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K+Hnov14	37299	yg64e08.r1	165015	5'
K+Hnov14	157854	yl10e04.r1	7, <b>251g07</b> (3, 34) (4)	5'
K+Hnov14	728558	zt79c08.r1	1787j15	5'
K+Hnov28	700757	zs48h03.r1	1715d6	5'
K+Hnov42	491748	zl08e07.s1	1170013	3'
K+Hnov42	491748	z108e07.r1	1170o13	5'
K+Hnov42	626699	zp82d06.r1	1522i12	5'
K+Hnov42	626699	zp82d06.s1	1522f12	3'
K+Hnov42	773611	zw51f10.r1	1904o20	5'
K+Hnov44	683888	zs01a05.s1	1671e9	3'
K+Hnov44	683888	zs01a05.r1	1671e9	5'
	K+Hnov14 K+Hnov14 K+Hnov28 K+Hnov42 K+Hnov42 K+Hnov42 K+Hnov42 K+Hnov42 K+Hnov44	K+Hnov14 157854 K+Hnov14 728558 K+Hnov28 700757 K+Hnov42 491748 K+Hnov42 491748 K+Hnov42 626699 K+Hnov42 626699 K+Hnov44 683888	K+Hnov14157854yl10e04.r1K+Hnov14728558zt79c08.r1K+Hnov28700757zs48h03.r1K+Hnov42491748zl08e07.s1K+Hnov42491748zl08e07.r1K+Hnov42626699zp82d06.r1K+Hnov42626699zp82d06.s1K+Hnov42773611zw51f10.r1K+Hnov44683888zs01a05.s1	K+Hnov14       157854       yl10e04.r1       251g07         K+Hnov14       728558       zt79c08.r1       1787j15         K+Hnov28       700757       zs48h03.r1       1715d6         K+Hnov42       491748       zl08e07.s1       1170o13         K+Hnov42       491748       zl08e07.r1       1170o13         K+Hnov42       626699       zp82d06.r1       1522f12         K+Hnov42       626699       zp82d06.s1       i522f12         K+Hnov42       773611       zw51f10.r1       1904o20         K+Hnov44       683888       zs01a05.s1       1671e9

## EXAMPLE 2: CHROMOSOMAL LOCALIZATION

Two primers were designed in the 3'-untranslated regions of each gene sequence to amplify a product across the Stanford G3 radiation hybrid map, or the Whitehead GB4 panel. The PCR data were submitted for automatic two-point analysis. Mapping data were correlated with cytoband information and comparisons with the OMIM human gene map data base were made. The following primers were made:

# K+Hnov1 on GB4 (SEQ ID NO:31) F: 5' TATCCACATCAATGGACAAGC 3' (SEQ ID NO:32) R: 5' TGCATAACTGGCTGGGTGTA 3' Results: 1.71 cR from D2S331, Cytogenetic location of 2g37

K+Hnov2 on G3
15 F: 5' GTCAGGTGACCGAGTTCA 3'
R: 5' GCTCCATCTCCAGATTCTTC 3'
Results: 0.0 cR from SHGC-1320, Cytogenetic location of 11q12

K+Hnov6 on GB4

20 (SEQ ID NO:33) F: 5' TGACATCACTGGATGAACTTGA 3'
(SEQ ID NO:34) R: 5' TGCCTGCAAAGTTTGAACAT 3'
Results: 5.23 cR from WI-5509, Cytogenetic location of 2p23

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K+Hnov9 on GB4
25 (SEQ ID NO:35) F: 5' TGACATCACTGGATGAACTTGA 3' (SEQ ID NO:36) R: 5' TGCCTGCAAAGTTTGAACAT 3'

Results: 1.21 cR from AFM200VC7, Cytogenetic location of 8q23

K+Hnov11 on GB4

(SEQ ID:NO:37) F: 5' ACCTGGTGGTATGGAAGCAT 3'

5 (SEQ ID NO:38) R: 5' TTTCTCCTGGCCTCTACCC 3'

Results: 2.43 cR from WI-6756, Cytogenetic location of 8q23

K+Hnov12 on G3

(SEQ ID NO:39) F: 5' TCCCTCTTGGGTGACCTTC 3'

(SEQ ID NO:40) R: 5' ATCTTTGTCAGCCACCAGCT 3' 10

Results: 7.45 cR from SHGC-32925, Cytogenetic location of Xp21

K+Hnov14 on GB4

(SEQ ID NO:41) F: 5' AGGTGTGCTGCCATCTGCTGTTCG3'

15 (SEQ ID NO:42) R: 5' AGCCTATCCTCTGAGAGTCAGG

Results:

7.69 cR from WI-7107, Cytogenetic location of 12q14

J. 10 1 18 18 18

K+Hnov28 on GB4

(SEQ ID NO:43) F: 5' AAGCAGAGTACTCATGATGCC 3'

20 (SEQ ID NO:44) R: 5' TCTGGTAGACAGTACAGTGG 3'

Results: 35.38 cR from WI-9695, Cytogenetic location of 3q29

K+Hnov42 on G3

(SEQ ID NO:45) F: 5' CATTTGGCTGGTCCAAGATG 3' Common Common

(SEQ ID NO:46) R: 5' AGTCATTGGTAGGGAGGTAC 3' 25

Results: 7.45 cR from SHGC-32925, Cytogenetic location of Xp21

K+Hnov44 on G3

(SEQ ID NO:47) F: 5' CATGCTTCTACAGTCCAGCC 3'

30 (SEQ ID NO:48) R: 5' GGTCCTCAGTTGCAGAAATC 3'

Results: 7.45 cR from SHGC-32925, Cytogenetic location of Xp21

Map positions for K+Hnov15 and K+Hnov27 were obtained from public databases. K+Hnov2 and K+Hnov4 have not been mapped.

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## EXAMPLE 3: EXPRESSION ANALYSIS

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RT-PCR was utilized to characterize the expression pattern of the novel ion channels. This approach used RNA from 30 different tissues to generate first strand cDNA. Total RNA was purchased (Clontech, Invitrogen) and used to synthesize first strand cDNA using M-MLV reverse transcriptase and the supplied buffer (Gibco-BRL). The 20 µl reaction contained 5 µg total RNA, 100 ng of random primers, 10 mM DTT,

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0.5 mM each dNTP, and an RNAse inhibitor (Gibco-BRL). Identical reactions were set up without reverse transcriptase to control for DNA contamination in the RNA samples. The synthesis reaction proceeded for 1 hour at 37°C followed by 10 minutes at 95°C. These cDNAs, along with control cDNA synthesis reactions without reverse transcriptase, were diluted 1.5 and 2 μl of each sample were arrayed into 96-well trays, dried, and resuspended in PCR buffer prior to PCR amplification. The cDNAs were tested with primers with defined expression patterns to verify the presence of amplifiable cDNA from each tissue. Gene-specific primers were used to amplify the cDNAs in 20 μl PCR reactions with standard conditions, 2.5 mM MgCl<sub>2</sub>, Taq Gold, and an appropriate annealing temperature.

This approach provides for relatively high-throughput analysis of gene expression in a large set of tissues in a cost-efficient manner and provides qualitative analysis of gene expression only. Modifications can be employed, such as the use of internal control primers, limited cycling parameters, and dilution series to convert this to a quantitative experiment.

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Testis	•	•	7	•	•	7	-	•	•	•	•	$\dashv$	•
Stomach	•	•	-				$\dashv$		-		+	+	-
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A \*\* indicates expression in the tissue, a \*\* indicates no expression, and blank square indicates no data for that sample.

P. S. & W. G. 1983

# K+Hnov49 on Whitehead GB4 RH mapping panel:

Primer 1 (SEQ ID NO:5): 5' - CATAGCCATAGGTGAGGACT - 3'

Primer 2: (SEQ ID N:6) 5" - GAGAGGAAAACAGTCTGGGC - 3"

5 Results: Cytogenetic location 1q41, 4.6cR from framework marker D1S217

# K+Hnov59 on Whitehead GB4 RH mapping panel

Primer 1 (SEQ ID NO:7): 5' - GGACATCGAACTAAGACCTG - 3'

Primer 2 (SEQ ID NO:8): 5' - TCCCATGCCATTCAGATCTG - 3'

10 Results: Cytogenetic location 19q13.2, 8.34cr from framework marker D19S425

## **EXPRESSION ANALYSIS OF K+HNOV49**

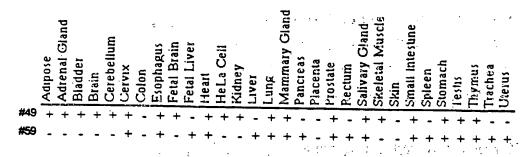
A probe was created from a fragment corresponding to nucleotides 50 to 1284 of SEQ ID NO:83 (K+Hnov49) and purified DNA fragment was labeled with [³²P]dCTP (Amersham) by the random primer method. Adult human Multiple Tissue Northern (MTM™) Blots (Clontech) were hybridized with the [³²P]-labeled fragment in ExpressHyb™ solution (Clontech) for four hours, washed to a final stringency of 0.1xSSC, 0.1% SDS at 65°C and subjected to autoradiography for 24 hours.

Analysis revealed that K+Hnov49 is expressed as an approximately 4.2kb mRNA. Expression levels of K+Hnov49 are high in brain and liver and low in kidney tissues. No mRNA was detectable on these Northern blots for heart, skeletal muscle, colon, thymus, spleen, small intestine, placenta, lung or peripheral blood leukocytes indicating either a very low level of expression or that it is not expressed in these tissues. Expression analysis was also carried out by RT-PCR across an extended series of tissues. The results of these analyses are shown in Table 4. Primer pairs used for amplification of K+Hnov49 and 59 are the same as those used for RH mapping as indicated above.

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### WHAT IS CLAIMED IS:

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- 1. An isolated nucleic acid encoding a mammalian K+Hnov protein.
- 2. An isolated nucleic acid according to Claim 1, wherein said K+Hnov protein has the amino acid sequence of SEQ ID NO:2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 25, 27, 30, 81 or 83.
- 3. An isolated nucleic acid according to Claim 1, wherein said K+Hnov protein has an amino acid sequence that is substantially identical to the amino acid sequence of SEQ ID NO:2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 25, 27, 30, 81 or 83.
- 4. An isolated nucleic acid according to Claim 1 wherein the nucleotide sequence of said nucleic acid is SEQ ID NO:1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 15 22, 23, 24, 26, 28, 29, 80 or 82.
  - 5. An isolated nucleic acid that hybridizes under stringent conditions to a nucleic acid sequence of claim 4.
  - 6. An expression cassette comprising a transcriptional initiation region functional in an expression host, a nucleic acid having a sequence of the isolated nucleic acid according to Claim 1 under the transcriptional regulation of said transcriptional initiation region, and a transcriptional termination region functional in said expression host.

7. A cell comprising an expression cassette according to Claim 6 as part of an extrachromosomal element or integrated into the genome of a host cell as a result of introduction of said expression cassette into said host cell, and the

cellular progeny of said host cell.

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8. A method for producing mammalian K+Hnov protein, said method comprising:

growing a cell according to Claim 7, whereby said mammalian K+Hnov protein is expressed; and

isolating said K+Hnov protein free of other proteins.

- 9. A purified polypeptide composition comprising at least 50 weight % of the protein present as a K+Hnov protein or a fragment thereof.
- 10. A monoclonal antibody binding specifically to a K+Hnov protein.
  - 11. A non-human transgenic animal model for K+Hnov gene function wherein said transgenic animal comprises an introduced alteration in a K+Hnov gene.

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- 12. The animal model of claim 11, wherein said animal is heterozygous for said introduced alteration.
- 13. The animal model of claim 12, wherein said animal is homozygous20 for said introduced alteration.
  - 14. The animal model of claim 12, wherein said introduced alteration is a knockout of endogenous K+Hnov gene expression.

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<221 <222 <223 <400 attaaaatta taaaggctga  aat tgc aa Asn Cys Ly 5	> CDS > (103)(1180) > K+Hnov1 > 1 > tetgateaaa aaggeaga cccagcaaaa gaactgaa a gtt att get eet ee s Val Ile Ala Pro Le	act ctgtaaattt (gaa atacagcctg a tc cta agt caa a au Leu Ser Gln )	ccttaagacc taccttgg ag atg gac agc agt Met Asp Ser Ser 1 aga tac cgg agg atg Arg Tyr Arg Arg Met 20	114
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<221 <222 <223 <400 attaaaatta taaaggctga  aat tgc aa Asn Cys Ly 5	> CDS > (103)(1180) > K+Hnov1 > 1 - tetgateaaa aaggeaga - cecageaaaa gaactgaa a gtt att get eet ee s Val Ile Ala Pro La 10 g gat gge eac age aa	act ctgtaaattt gaa atacagcctg atc cta agt caa aeu Leu Ser Gln 1	ccttaagacc taccttggo ag atg gac agc agt Met Asp Ser Ser 1 aga tac cgg agg atg Arg Tyr Arg Arg Met 20	114
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<pre>&lt;221 &lt;222 &lt;223 &lt;400 attaaaatta taaaggctga  aat tgc aa Asn Cys Ly 5 gtc acc aa Val Thr Ly</pre>	> CDS > (103)(1180) > K+Hnov1 > 1 - tetgateaaa aaggeaga - cecageaaaa gaactgaa a gtt att get eet ee s Val Ile Ala Pro Le 10 g gat gge eac age ac s Asp Gly His Ser Th	act ctgtaaattt gaa atacagcctg atc cta agt caa atacagcctg atc cta agt caa atg caa atg car Leu Gln Met 1	ccttaagacc taccttggdag atg gac agc agt Met Asp Ser Ser 1 aga tac cgg agg atg Arg Tyr Arg Arg Met 20 gat ggc gct caa aga Asp Gly Ala Gln Arg 35	114 162 210
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Leu	Gln	H1S	Glu	Asn	<b>Pro</b>	Ser	His	Phe	Glu	:Leu	Val.	.Val	Phe	Leu	Ser	
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ggag agto gag Glu 5	9c9 41a	G1u 355 210> 211> 212> 221> 222> 23> 400> gc a yaa c	Thr 3 1927 DNA H. s CDS (105 K+Hn 3 agege caage	apie ov4 ettet ettec	ens (190 et ate g to ate Ile 10	gato caaa atc Ile	agct acat atc Ile	egg gtc aac Asn	tgtg:tgaa	ggc ggc Gly 15	ggc	etect atg Met 1 acg Thr	ac o gco Ala cga Arg	cgcgc caac Lys cat His	gag Glu 20	1 60 116
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ggag agto gag Glu 5	<pre>&lt;2 &lt;2 &lt;4 geogg Ala tac</pre>	G1u 355 210> 211> 212> 221> 222> 23> 400> 450 5er	Thr  3 1927 DNA H. s  CDS (105 K+Hn 3 agegee caage	apie ov4 ttct ctcc	ens (190 ct atc atc Ile 10 ctg	gato caaa atc Ile	agct acat atc Ile	e cgg gtc aac Asn	gtg Val	ggc Gly 15	ggc ggag	etect atg Met 1 acg Thr	ac o gco Ala cga Arg	egege e aac Lys cat His gcc Ala	gag Glu 20	1 60 116
ggag agto gag Glu 5 acc	<pre> <pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre>	G1u 355 210> 211> 212> 221> 222> 23> 400> 400> 400> 400> 400> 400> 400> 40	Thr  3 1927 DNA H. s  CDS (105 K+Hn 3 agcgc caagc	apie ov4 ettet ctcc aag Lys acc Thr	ens (190 atc Ile 10 ctg Leu	gato caaa atc Ile cgc Arg	agct acat atc Ile acc Thr	e cgg gtc aac Asn cta Leu	gtg Val ccg Pro	ggc Gly 15 gga Gly	ctcc ggag ggc Gly acc Thr	acg Thr	cga Arg ctc Leu	cgcgc caac Lys cat His gcc Ala 35	gag gag Glu 20 tgg Trp	164 212
ggag agto gag Glu 5 acc Thr	<pre> <pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre></pre>	G1u 355 210> 211> 212> 221> 223> 200> 221> 223> 23c 24c 25c 26c 27c 27c 27c 27c 27c 27c 27c 27c 27c 27	Thr  3 1927 DNA H. s  CDS (105 K+Hn 3 agcgc caagc	apie ov4 ttct ctcc aag Lys acc Thr 25 gac	ens (190 atc Ile 10 ctg Leu	gato caaa atc Ile cgc Arg	agct acat atc Ile acc Thr	aac Asn cta Leu	gtg Val ccg Pro 30	ggc Gly 15 gga Gly	ctcc ggag ggc Gly acc Thr	acg Thr cgc Arg	ac o gcc Ala cga Arg ctc Leu	cgcgc caac Lys cat His gcc Ala 35	gag Glu 20 tgg Trp	1 60 116

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Asp	70	cac His	Pro	Gly	Val	Phe 75	Ala	Tyr	Val	Leu	Asn 80	Tyr	Tyr	Arg	Thr	356	5
85 GIÀ	гуs	ctg Leu	His	Cys	Pro 90	Ala	Asp	Val	Cys	Gly 95	Pro	Leu	Phe	Glu	Glu 100	404	ì
GIu.	Leu	gcc Ala	Phe	Trp 105	Gly	Ile	Asp	Glu	Thr 110	Asp	Val	Glu	Pro	Cys 115	Суѕ	452	2
Trp	Met	acc Thr	120	Arg	Gln	His	Arg	Asp 125	Ala	Glu	Glu	Ala	Leu 130	Asp	Ile	500	)
Pne	GIU	acc Thr 135	Pro	Asp	Leu	Ile	Gly 140	Gly	Asp	Pro	Gly	Asp 145	Asp	Glu	Asp	548	}
Leu	150	gcc Ala	Lys	Arg	Leu	Gly 155	Ile	Glu	Asp	Ala	Ala 160	Gly	Leu	Gly	Gly	596	i
165	Asp	Gly	Lys	Ser	Gly 170	Arg	Trp	Arg	Arg	Leu 175	Gln	Pro	Arg	Met	Trp 180	644	:
Ala	Leu	ttc Phe	Glu	185	Pro	Tyr	Ser	Ser	Arg 190	Ala	Ala	Arg	Phe	Ile 195	Ala	692	
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A1a 245	Leu	acg Thr	Tyr	Val	Glu 250	Gly	Val	Cys	Val	Val 255	Trp	Phe	Thr	Phe	Glu 260	884	
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val Inr Ser	Pro Tyr As	n Ser Pro Cy	s Pro Leu Arg	Arg Ser Arg	tct 1892 Ser
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	H. sapiens				
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~ 35 ~	•	40		4.5	
Asp GIV GIV	GIY VAI GIY	Ser Ser Gly	Ser Ser Gly	Gly Gly Gly	Cys
Glu Phe Phe	Phe Asp Arg	His Pro Gly	/ Val Phe Ala	Tyr Val Leu	Asn
65 Tyr Tyr Arg	Thr Gly Lys	Leu His Cvs	75 Pro Ala Asp	Val Cvs Glv	80 Pro
•	85		90	· 95	_
red bue GIG	GIU GIU Lei	Ala Phe Tri 105	Gly Ile Asp	Glu Thr Asp	Val
Glu Pro Cys	Cys Trp Met		Gln His Arg	Asp Ala Glu	Glu
115 Ala Leu Asp	Ile Phe Glu	120 Thr Pro Ast	Leu Ile Gly	Gly Asp Pro	Glv
130		135	140	* *	
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285

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 Arg Ile Phe Lys Leu Thr Arg His Phe Val Gly Leu Arg Val Leu Gly
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His Thr Leu Arg Ala Ser Thr Asn Glu Phe Leu Leu Leu Ile Ile Phe
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Leu Ala Leu Gly Val Leu Ile Phe Ala Thr Met Ile Tyr Tyr Ala Glu
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Arg Val Gly Ala Gln Pro Asn Asp Pro Ser Ala Ser Glu His Thr Gln
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Phe Lys Asn Ile Pro Ile Gly Phe Trp Trp Ala Val Val Thr Met Thr
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Thr Leu Gly Tyr Gly Asp Met Tyr Pro Gln Thr Trp Ser Gly Met Leu
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Val Gly Ala Leu Cys Ala Leu Ala Gly Val Leu Thr Ile Ala Met Pro
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Val Pro Val Ile Val Asn Asn Phe Gly Met Tyr Tyr Ser Leu Ala Met
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Ala Lys Gln Lys Leu Pro Arg Lys Arg Lys Lys His Ile Pro Pro Ala
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Pro Gln Ala Ser Ser Pro Thr Phe Cys Lys Thr Glu Leu Asn Met Ala
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Cys Asn Ser Thr Gln Ser Asp Thr Cys Leu Gly Lys Asp Asn Arg Leu
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Leu Glu His Asn Arg Ser Val Leu Ser Gly Asp Asp Ser Thr Gly Ser
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Glu Pro Pro Leu Ser Pro Pro Glu Arg Leu Pro Ile Arg Arg Ser Ser
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Thr Arg Asp Lys Asn Arg Arg Gly Glu Thr Cys Phe Leu Leu Thr Thr
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Gly Asp Tyr Thr Cys Ala Ser Asp Gly Gly Ile Arg Lys Gly Tyr Glu
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                                         555
Lys Ser Arg Ser Leu Asn Asn Ile Ala Gly Leu Ala Gly Asn Ala Leu
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gagaacagga ttcttccctt ctttttggcc accaaatgcc tatgtgcacc acacattcca
                                                                      180
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                                                                      240
ggcagagcgt gtggcatete caceteaagg gtgcagcetg atetteetet tetecettge
                                                                      300
cagccagcac tetgeettet gtatecace atg gtg ttt ggt gag ttt tte cat
Met Val Phe Gly Glu Phe Phe His
                                                                      353
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ε

5 ) 3 4 (1 ) (1 ) (1 ) (1 )

cgc	cct	gga	caa	gac	gag	gaa	ctt	gtc	aac	cta	aat	ata	aaa	gac	+++	401
Arg	Pro	Gly	Gln	Asp	Glu	Glu	Leu	Val	Asn	Leu	Asn	Val	Glv	Glv	Dhe	401
_	10	-		•		15					20.		<b>U T Y</b>	GLY	2116	
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aag	cag	tct	gtt	gac	caa	agc	acc	cte	cta	caa	ttt	cct	cac	acc	acra	449
Lys	Gln	Ser	Val	Asp	Gln	Ser	Thr	Leu	Leu	Arg	Phe	Pro	Hie	Thr	Ara:	447
25				•	30					35				****	40	
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cta	aaa	aag	ctg	ctt	act	tac	cat	tot	maa				a+~	~~~		407
Leu	Glv	Lvs	Leu	Leu	Thr	Cvs	Hig	Ser	Glu	Glu	712	Tla	Lou	gag	Tan	497
	1	-1-		45		-75	****		. 50	GIU	ALA	TIE	Leu			
	4	• • •								:				. 55		
tat	gat	gat	tac	agt	ata	acc	gat	220	caa	tac	tac		~~+			545
Cvs	Asp	Asp	Tyr	Ser	Val	Δla	Agn	Tare	Glu	7.~	Tree	Dho	yac Nan	299	aac	545
-7-			60			712.4	LSP	65	014	TYL	TAT	FILE		Arg	Asn	
-			•			•		0.5	. *				70			
CCC	fice	tta	ttc	arra	tät	att				+						
Pro	Ser	Len	Phe	Ara	Tur	Val	Len	Agn	Pho	The	The	acg	999	aag	ctg	593
	001	75		y	- 7 -	447		Wall	FILE	TAT	TÄL		GIA	гÀг	Leu	
					'		. 80	-	. 2 .	111	•	85			٠.	
car	atc	ato	gag	a si a	CEG	+ 44	a+ a	++-	+ 42		<u>:</u>	: :	·.		**	
Hig	Val	Met	Glu	Glu	Leu	Cyc	y ca	Dho	Coa	Dho	cge	cag	gag	acc	gag	641
	90	1-16-0	GIU	GIU	neu		Val	Pne	ser	Pne		Gin	GIU	Ile	Glu	
	30	•	. ::	. •		95	٠.٠				100			•		
tac	+00	-	atc		<b>~</b> 3~										'	
Tur	T	Clu	Tlo	aac aac	gag	Tou	בבכ	att	gat	TCE	tgc	tgc	agc	aat	cgc	689
105	пр	GIY	Ile	ASII		Leu	Pne	TTE	Asp		Cys	Cys	Ser	Asn	_	
105	. 3				110		*		•	115	;			•	120	
t 2.0	مدخ						11_	<u> </u>				42.6			,	
The state	Cay	gaa	cgc	aag	gag	gaa	aac	cac	gag	aag	gac	tgg	gac	cag	aaa	737
TYL	GIII	GIU	Arg		GIU	GIU	Asn	Hls		rys	Asp	Trp	Asp		Lys	
				125					130		t			1,35	7	
						_:_'					<u>`</u>			••		
age	Cat	gat	gtg	agt	acc	gac	FCC	tcg	בככ	gaa	gag	tcg	tct	ctg	ttt	785
Ser	HIS	Asp	Val	ser	Thr	Asp	ser		Pne	Glu	Glu	Ser			Phe	•
			140			100		145	,		:	-	150			
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Glu	Tue	gag	ctg	gag	aag	בבב	gac	aca	ctg	cga	בכב	aar	cag	ctc	cgg	833
Giu	rAs_	155	Leu	GIU	гÀв	Pne		Thr	Leu	Arg		_		Leu	Arg	
		133	•				160	5. (	* .		-	165				
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aag	aaa	acc	tgg	att	aga	atg	gag	aat	cca	aca	tac	tgc	ctg	tcc	gct	881
гλа	Lys	TIE	Trp	TTE	Arg	met	GIU	Asn	Pro	Ala	Tyr	Cys	Leu	Ser	Ala	•
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						<u>.</u>	. :						•	**	•	
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			Val			Met	Ser	Glu							Glu	
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gtg	gat	gat	ccg	gtg	ctg	gaa	gga	gtg	gag	atc	gcg	tgc	att	gcc	tgg	1025
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Phe	Thr	Gly	Glu	Leu	Ala	Val	Arg	Leu	Ala	Ala	Ala	Pro	Cys	Gln	Lys	
															٠ <u>.</u> -	
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gtg Val 345	gag Glu	aaa Lys	gat Asp	gac	cac His 350	aca Thr	tcc Ser	agc Ser	ctc Leu	acc Thr 355	agc Ser	atc Ile	ccc Pro	atc Ile	tgc Cys 360	1409
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cac His	ccg Pro	gtc Val	acc Thr 380	ttg Leu	gcg Ala	gga Gly	aag Lys	ctc Leu 385	atc Ile	gcc Ala	agc Ser	aca Thr	tgc Cys 390	atc Ile	atc Ile	1505
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agg Arg	gat Asp	ata Ile	tat Tyr	gca Ala 445	Gln	Arg	atg Met	cac His	gcc Ala 450	ttc Phe	att Ile	acc Thr	agt Ser	ctc Leu 455	tct Ser	1697
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490

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                                     395
Pro Ile Thr Ile Ile Phe Asn Lys Phe Ser Lys Tyr Tyr Gln Lys Gln
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Lys Asp Ile Asp Val Asp Gln Cys Ser Glu Asp Ala Pro Glu Lys Cys
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His Ala Phe Ile Thr Ser Leu Ser Ser Val Gly Ile Val Val Ser Asp
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                                                                    420
aagetteegg egtgteecea aetttgtgge geeeteagge egeggegaet gggttagag
                                                                    479
atg cct tee age gge aga geg etg etg gae teg eeg etg gae age gge
                                                                    527
Met Pro Ser Ser Gly Arg Ala Leu Leu Asp Ser Pro Leu Asp Ser Gly
tee etg ace tee etg gae tet agt gte tte tge age gag ggt gaa ggg
                                                                    575
Ser Leu Thr Ser Leu Asp Ser Ser Val Phe Cys Ser Glu Gly Glu Gly
gag ccc ttg gcg ctc ggg gac tgc ttc acg gtc aac gtg ggc ggc agc
                                                                    623
Glu Pro Leu Ala Leu Gly Asp Cys Phe Thr Val Asn Val Gly Gly Ser
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ege the gtg etc teg cag cag geg etg tee tge tte eeg cac acg ege
                                                                    671
Arg Phe Val Leu Ser Gln Gln Ala Leu Ser Cys Phe Pro His Thr Arg 50 55 60
ctt ggc aag ctg gcc gtg gtg gct tcc tac cgc cgc ccc ggg gcc
                                                                    719
Leu Gly Lys Leu Ala Val Val Val Ala Ser Tyr Arg Arg Pro Gly Ala
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ctg gcc gcc gtg ccc agc cct ctg gag ctt tgc gac gat gcc aac ccc
Leu Ala Ala Val Pro Ser Pro Leu Glu Leu Cys Asp Asp Ala Asn Pro
                                   90
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155 . 310 . 51

12 5 66

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					٠.								110	• • •	\$ 1.5	
gtc	ctg	cac	tac	tac	cgc	acc	990	cac	cta	cat	atc	ato	gag	Cac	cta	863
Val	Leu	His	Tyr	Tyr	Arg	Thr	Gly	Arg	Leu	His	Val	Met	Glu	Glr	T.011	863
		115		_	_		120	_				125				
	, f		:		•	٠,٠				:						
tgc	gcg	ctc	tcc	ttc	ctg	cag	gag	atc	cag	tac	tgg	ggc	atc	gat	gag	911
Cys	ALA	Leu	Ser	Phe	Leu	Gln	Glu	Ile	Gln	Tyr	Trp	Gly	Ile	Asp	Glu	
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Tou	agc	Tle	gat	C,CC	tgc	tgc	agg	gac	aga	tac	ttc	aga	agg	aaa	ġag	959
145	Ser	116	ASD	ser	150	Cys	Arg	Asp	Arg		Phe	Arg	Arg	Lys		
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ctq	agt	gaa	act	tta	gac	ttc	aag	aaσ	gac	202		gad.	~~~			1007
Leu	Ser	Glu	Thr	Leu	Asp	Phe	Lvs	Lvs	Asp	Thr	Glu	Asn	Gla	Glu	Ser	1007
				165			-1-	-7-	170		014	vab	GIII	175	Ser	
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caa	cat	gag	agt	gaa	cag	gac	ttc	tcc	caa	gga	cct	tat	ccc	act	att	1055
Gln	His	Glu	Ser	Glu	Gln	Asp	Phe	Ser	Gln	Gly	Pro	Cys	Pro	Thr	Val	
			180					185		_		•	190			
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cgc.	cag	aag	CTC.	tgg	aat	atc	ctg	gag	aaa	cct	gga	tct	tcc	aca	gct	1103
Arg	GIN	195	ren.	Trp	ASD	Ile		Glu	Lys	Pro	Gly		Ser	Thr	Ala	
		173			, ,	4	200					205				
acc	cat	atc	ተተ	ממכ	a+ċ	acc	+00									
Ala	Arg	Ile	Phe	Glv	Val	Ile	Ser	Tle	Tla	Dha	geg	grg	gcg	CCC	atc	1151
	210			,		215	561	116	116	FILE	220	Val	vai	ser	116	
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Ile	Asn	Met	Ala	Leu	Met	Ser	Ala	Glu	Leu	Ser	Trp	Leu	Asp	Leu	Gln	
225					230					235		,	_		240	
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ctg	ctg	gaa	atc	ctg	gag	tat	gtg	tgc	att	agc	tgg	ttc	acc	999	gag	1247
nea	Letu	GIU	TTE	245	GIU	Tyr	Val	Cys		Ser	Trp	Phe	Thr		Glu	
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ttt	atc	ctc			cta	tgt	ata	-	<i>~</i> 2 <i>~</i>	~~~			2		_ :_ :	1005
Phe	Val	Leu	Ara	Phe	Leu	Cys	Val	Ara	Agn	Ara	Cve	Ara	Dhe	Cta	aga	1295
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aag	gtg	cca	aac.	atc	ata	gac	ctċ	ctt	gcc	atc	ttq	ccc	ttc	tac	atc	1343
Lys	Val	Pro	Asn .	Ile	Ile	qaA	Leu	Leu	Ala	Ile	Leu"	Pro	Phe	Tyr	Ile	
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act.	ctt	ctg	gta	gag	agc	cta	agt	<b>ā</b> 33	agc	cag	acc	acg	cag	gag	cţg	1391
Int	neu	Leu 1	val	GIU.	ser	Leu	Ser	Gly	Ser	Gln	Thr	Thr	Gln	Glu	Leu	
						295			. 1		300	•				
			aaa	caa				~+-						22	_£	
Gl u	Agn	Val	G] v	Ara	Tla	gtc Val	cag	yeg '	Leg	agg	ctg	CCC	agg	gct	ctg	1439
3.05	, ,		<b></b> y		310.	AGT		va.	-cu	315	neń	neu	wid		120	
						17 BE		· . •	s' .	J_J .					J2U	
					ggc	aga	cat	tee	aca	gga	tta	čać	tec	ctt	ggg	1487
Arg	Met	Leu	Lys	Leu	Gly	Arg	His	Ser	Thr	Glv	Leu	Ara	Ser	Leu	Glv	_TU/
. 17		•		325	₹.			· ,	330	4	<del>-</del> -	, <b>3</b>	٦٠٠,	3,35		
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48-23 A. Same

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3080

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Ile Leu Val Leu Ala Leu Pro Ile Ala Ile Ile Asn Asp Arg Phe Ser

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	450				Leu	455					460	•		-		
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Asp	Phe	Trp		,	•				•	·				•••	-f	
		210>				; <i>.</i>				٠.٠		٠	: :	<i>:</i> .	Ť	
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			DNA B.	sapi	ens .				,			٠.	٠			
		220>				, .		; .	٠.		, .					
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Ala	Asp	Ser	Gly	Glu	Tyr	Phe	Phe	Asp	Arg	Asp	Pro	Asp	Met	Phe	Arg	532
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EDC 717

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Ile Ile Ile Phe Ala Thr Val Met Phe Tyr Ala Glu Lys Gly Thr Asn 340 345 350 350 Lys Thr Asn Phe Thr Ser Ile Pro Ala Ala Phe Trp Tyr Thr Ile Val Thr Met Thr Leu Gly Tyr Gly Asp Met Val Pro Ser Thr Ile Ala 370 380 Gly Lys Ile Phe Gly Ser Ile Cys Ser Leu Ser Gly Val Leu Val Ile 390 395 400 Ala Leu Pro Val Pro Val Ile Val Ser Asn Phe Ser Arg Ile Tyr His 405 410 Gln Asn Gln Arg Ala Asp Lys Arg Arg Ala Gln Gln Lys Val Arg Leu 420 425 Ala Arg Ile Arg Leu Ala Lys Ser Gly Thr Thr Asn Ala Phe Leu Gln 440 445 Tyr Lys Gln Asn Gly Gly Leu Glu Asp Ser Gly Ser Gly Glu Gln 455 460 Ala Leu Cys Val Arg Asn Arg Ser Ala Phe Glu Gln Gln His His His 465 470 470 470 475 Leu Leu His Cys Leu Glu Lys Thr Thr Cys His Glu Phe Thr Asp Glu 485 490 Leu Thr Phe Ser Glu Ala Leu Gly Ala Val Ser Pro Gly Gly Arg Thr 505 Ser Arg Ser Thr Ser Val Ser Ser Gln Pro Val Gly Pro Gly Ser Leu 520 Leu Ser Ser Cys Cys Pro Arg Arg Ala Lys Arg Arg Ala Ile Arg Leu 530 535 540 Ala Asn Ser Thr Ala Ser Val Ser Arg Gly Ser Met Gln Glu Leu Asp 550 555 Met Leu Ala Gly Leu Arg Arg Ser His Ala Pro Gln Ser Arg Ser Ser 565 570 Leu Asn Ala Lys Pro His Asp Ser Leu Asp Leu Asn Cys Asp Ser Arg 580 585 Asp Phe Val Ala Ala Ile Ile Ser Ile Pro Thr Pro Pro Ala Asn Thr 595 600 Pro Asp Glu Ser Gln Pro Ser Ser Pro Gly Gly Gly Arg Ala Gly 610 ... 615 620 Ser Thr Leu Arg Asn Ser Ser Leu Gly Thr Pro Cys Leu Phe Pro Glu 630 635 640 Thr Val Lys Ile Ser Ser

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cac age gat gea	ctt cat ttt atc a a	gtaattacc tgtgtcacga	1177
His Ser Asp Ala		ing the state of t	
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Met Glu Arg Lys  1 Lys His Asn Ser 20 Thr Leu Met Thr 35 Gln Thr Leu Thr	Ile Asn Arg Arg Glu 5 Leu Glu Asp Thr Asp 25 Leu Asn Val Gly Gly 40 Lys Tyr Pro Asp Thr	Lys Glu Lys Glu Tyr Glu Gly 10 15 Gln Gly Lys Asn Cys Lys Ser 30 Tyr Leu Tyr Ile Thr Gln Lys 45 Phe Leu Glu Gly Ile Val Asn	,
Met Glu Arg Lys  1 Lys His Asn Ser 20 Thr Leu Met Thr 35 Gln Thr Leu Thr	Ile Asn Arg Arg Glu 5 Leu Glu Asp Thr Asp 25 Leu Asn Val Gly Gly 40 Lys Tyr Pro Asp Thr 55	Lys Glu Lys Glu Tyr Glu Gly 10 15 Gln Gly Lys Asn Cys Lys Ser 30 Tyr Leu Tyr Ile Thr Gln Lys 45 Phe Leu Glu Gly Ile Val Asn 60	
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Ala Ser Pro Leu Xaa Asn Gln Gly Ile Pro Thr Pro Ala Gln Leu Thr
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Lys Ser Asn Ala Pro Val His Ile Asp Val Gly Gly His Met Tyr Thr
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Leu Phe Asp Gly Thr Glu Pro Ile Val Leu Asp Ser Leu Lys Gln His
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Tyr Phe Ile Asp Arg Asp Gly Gln Met Phe Arg Tyr Ile Leu Asn Phe
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Leu Arg Thr Ser Lys Leu Leu Ile Pro Asp Asp Phe Lys Asp Tyr Thr
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ccc tgt gag tgc ctc g Pro Cys Glu Cys Leu V 140	tc gtg cgt gtg gcc cca al Val Arg Val Ala Pro 145	gac ctc gga gaa agg 783 Asp Leu Gly Glu Arg 150
Ile Thr Leu Ser Gly A	ac aaa tcc ttg ata gaa sp Lys Ser Leu Ile Glu 60 165	gaa gta ttt cca gag 833 Glu Val Phe Pro Glu 170
Ile Gly Asp Val Met C	gt aac tct gtc aat gca ys Asn Ser Val Asn Ala 180	Gly Trp Asn His Asp 185
Ser Thr His Val Ile Am	gg ttt cca cta aat ggc rg Phe Pro Leu Asn Gly 195	Tyr Cys His Leu Asn 200
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Pro Ile Val Leu Asp Ser Leu Lys Gln His Tyr Phe Ile Asp Arg Asp
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Cys ::.	Leu	Leu		Val	Leu	Ser	Arg		Arg	Ala	Trp	Val		Val	His	

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Tri	Gli		ı Ser	Pro	בוג י	Δ ***	40 - 11	. או	T			45				
	50					55					60					
00					70.					75					Leu	
Trp	Gly	/ Let	ı Gln	Gly 85	Asp	Суз	Ser	Leu	Leu 90	Gly	Ala	Val	Tyr	Phe	Cys	
Phe	s Ser	Ser	Leu 100	Ser	Thr	Ile	Gly	Leu	Glu			Leu			Arg	
Gly	Arg	Ser 115	Leu		Pro	Val	Ile	105 Tyr	His	Leu	Gly	Gln	110 Leu	Ala	Leu	
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CCC	egee.	agg (	cacac	acad	eg et	iacad	rcega	e cgo	tgtt	ccc	teeg	gette	ca g	gtgt	agcgc tgg	
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فينافر

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                                   135
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#### <213> H. sapiens

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Val Gly Phe Gly Asn Val Ser Ala Asn Thr Asp Thr Glu Lys Ile Phe 470 475 Ser Ile Cys Thr Met Leu Ile Gly Ala Leu Met His Ala Val Val Phe 485 490 Gly Asn Val Thr Ala Ile Ile Gln Arg Met Tyr Ala Arg Arg Phe Leu 505 Tyr His Ser Arg Thr Arg Asp Gln Arg Asp Tyr Ile Arg Ile His ary 520 525 Ile Pro Lys Pro Leu Lys Gln Arg Met Leu Glu Tyr Phe Gln Ala Thr 535 540 Trp Ala Val Asn Asn Gly Ile Asp Thr Thr Glu Leu Leu Gln Ser Leu 550 555 Pro Asp Glu Leu Arg Ala Asp Ile Ala Met His Leu His Lys Glu Val 565 570 Leu Gln Leu Pro Leu Phe Glu Ala Ala Ser Arg Gly Cys Leu Arg Ala 580 585 Leu Ser Leu Ala Leu Arg Pro Ala Phe Cys Thr Pro Gly Glu Tyr Leu 600 Ile His Gln Gly Asp Ala Leu Gln Ala Leu Tyr Phe Val Cys Ser Gly 615 620 Ser Met Glu Val Leu Lys Gly Gly Thr Val Leu Ala Ile Leu Gly Lys 630 635 Gly Asp Leu Ile Gly Cys Glu Leu Pro Arg Arg Glu Gln Val Val Lys 645 650 Ala Asn Ala Asp Val Lys Gly Leu Thr Tyr Cys Val Leu Gln Cys Leu 660 665 Gln Leu Ala Gly Leu His Asp Ser Leu Ala Leu Tyr Pro Glu Phe Ala 675 680 Pro Arg Phe Ser Arg Gly Leu Arg Gly Glu Leu Ser Tyr Asn Leu Gly 695 700 Ala Gly Gly Ser Ala Glu Val Asp Thr Ser Ser Leu Ser Gly Asp 710 715 Asn Thr Leu Met Ser Thr Leu Glu Glu Lys Glu Thr Asp Gly Glu Gin 725 730 Gly Pro Thr Val Ser Pro Ala Pro Ala Asp Glu Pro Ser Ser Pro Leu 740 745 Leu Ser Pro Gly Cys Thr Ser Ser Ser Ser Ala Ala Lys Leu Leu Ser 760 Pro Arg Arg Thr Ala Pro Arg Pro Arg Leu Gly Gly Arg Gly Arg Pro 775 780 Gly Arg Ala Gly Ala Leu Lys Ala Glu Ala Gly Pro Ser Ala Pro Pro 790 795 Arg Ala Leu Glu Gly Leu Arg Leu Pro Pro Met Pro Trp Asn Val Pro 805 810 Pro Asp Leu Ser Pro Arg Val Val Asp Gly Ile Glu Asp Gly Cys Gly 820 825 Ser Asp Gln Pro Lys Phe Ser Phe Arg Val Gly Gln Ser Gly Pro Glu 840 845 Cys Ser Ser Ser Pro Ser Pro Gly Pro Glu Ser Gly Leu Leu Thr Val .. 855 860 Pro His Gly Pro Ser Glu Ala Arg Asn Thr Asp Thr Leu Asp Lys Leu 870 875 Arg Gln Ala Val Thr Glu Leu Ser Glu Gln Val Leu Gln Met Arg Glu 885 890 Gly Leu Gln Ser Leu Arg Gln Ala Val Gln Leu Val Leu Ala Pro His 905 Arg Glu Gly Pro Cys Pro Arg Ala Ser Gly Glu Gly Pro Cys Pro Ala 915 920 925 Ser Thr Ser Gly Leu Leu Gln Pro Leu Cys Val Asp Thr Gly Ala Ser . 935 Ser Tyr Cys Leu Gln Pro Pro Ala Gly Ser Val Leu Ser Gly Thr Trp

945					950					955					960	
Pro	His	Pro	Arg	Pro 965	Gly	Pro	Pro	Pro	Leu 970	Met	Ala	Pro	Arg	Pro 975	Trp	
Gly	Pro	Pro	Ala 980	Ser	Gln	Ser	Ser			Pro	Arg	Ala		Ala	Phe	N
Trp:	Thr	Ser			Asp	Ser	Glu 100		Pro	Àla	Ser			Leu	Cys	
Ser	Glu 101	Pro	Ser	Thr	Pro	Ala 1019	Ser		Pro	Pro	Ser 1020		Glu	Gly	Ala	
Arg	Thr		Pro	Ala	Glu 103	Pro		Ser	Gln		Glu		Thr	Ser		
		Pro	Pro	Pro	Gly	Ser	Gly	Gly			Leu	Pro	Trp			
His	Ser	Leu	Glu	104! Met		Leu	Ile			) His	Gly	Ser	Gly	1059 Thr	Val	
	_	_,	1060					106					1070	כ		
GIn,	Trp			GIu	GIu	Gly		_	Val	•	•			•		
		1079	•				1080									
•		210>:	2.						•	•				•	•	
		210>: 211>		0			ι,			· ;	-				3	
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	<:	213>	н. я	sapi	ens		• •		*	*.	i .	, , ,	•	-	. :-	
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	٠.						•					•	. ,,	*;		
	< 4	100>	21													
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tttc	ctctq	gaa a	aatct	ttcaç	gt ct	ctta	agtto	cag	jatg	ggtt	ctct	atgo	ta o	gaat	acagg	: 60 120
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sagares e e e

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aag Lys 165	cac His	atg Met	atg Met	gac Asp	acc Thr 170	Arg	Asp	tgc Cys	cag Gln	gtt Val 175	Ser	ttt Phe	act Thr	ttt Phe	gga Gly 180	885
ccc Pro	tgt Cys	gat Asp	Tyr	cac His 185	Gln	gaa Glu	gtt Val	tct Ser	ctt Leu 190	agg Arg	gtc Val	cac His	ctg Leu	atg Met 195	gaa Glu	933
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	C+C		202		3.00		+	n.c.					:	<u> </u>		÷	
	Tau	The	The	ttg	The	7 = ~	m	CCG	gac	200	acg	CEE	gga	gct	atg	בנד -	507
	Deu	TILL	1111	Leu 30	1111	Arg	IÀI	PIO		ser	Mec	Leu	GIY		Met	Phe	
				30					35			9.5	•	40		1.	
	aaa	aaa.	gac.	ttc	dec	202	act	-	Ga C								555
	333	61 v	Acr.	Phe	Pro	Thr	300	Oya N==	gac	DWO.	Caa	ggc	aat	tac	בבב	att	555
	Gry	Gry	45		FIO	1111	AIG	50	MSP	PIO	GTÜ	GIY		Tyr	Pne	TIE	
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	gat	сста	gat	gga	cct	C++	÷÷c	CCT	* = <b>+</b>	ت جنہ	or o						
	Asp	Ara	Asn	Gly	Pro	Len	Phe	Ara	Tur	Val	Len	aac aac	Dha	Tou	aya	act The	603
		60				204	65	9	- 7 -	V 4 1	nea	70	FILE	Dea	Arg	1111	
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	Ser	Glu	Leu	Thr	Leu	Pro	Leu	Asp	Phe	Lvs	Glu	Phe	Asn	Leu	T.e.	Ara	651
	75					80				-,-	85		OP		<b>1</b> 100	90	
-												· . •				,,,	
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•	Leu	Ser		Thr	Arg.	Lys	Leu	Ser	Lys	Tyr	Ser	Asn	Pro	Val	Ala	Val	
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	202	~ ~ ~	+~~	636	~++	+00	+++	205		<b></b>							030
	۸×م	)ac	Cyc	cag Gln	y.1	505	Dho	The	Dho	gga	200	Con	gat	Tat.	cac	cag	939
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	ma a	att	tet	ctt	200	atc	Cac	cta	at~	(T22	***	2++	262			aa+	007
				Leu													987
		+ 0.1	x	190	~-4	• <b>a</b> I	****	~∈u	195	GIU	TÅT	TTE		200	GIII	Gry	
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Lys Thr Asp Asp
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                                                                                             125
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   Phe Thr Phe Gly Pro Cys Asp Tyr His Gln Glu Val Ser Leu Arg Val
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q a	tg'a	ga c	वव व	to a	cc ć	ta t	tc c	to a	ac a	ac a	ac c	CC 2	3~ 3	3533	ga aag	
- м	- A	ra A	V	- J T	h = T	- J - D	ho T	3	2	3	900	cc a	ay a	ac g	ga aag	229
1-1-	- -	-9 ~	-9 v	a1 1	III L	eu P	ne L	eu A	sn G	TA 2	er P	ro L	ys A	sn G	ly Lys	
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ata	~++	act	<b>~</b> + ~			_:			· .	1. "						
9.59	900	get	gta	Lat	gga	act	tta	CCC	gat	ttg	ctt	tct	gtg	gcc	agc	277
Val	Val	Ala	Val	Tyr	Gly	Thr	Leu	Ser	Asp	Leu	Leu	Ser	Val	Ala	Ser	
			20					25	-							
. •					٠.			. 20	;				, 30	•		
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Ser	LVS	Len	Glv	Tla	Lve	Δla	Th-	50-	77-3	T	3	Gly		330	994	223
	-,-		O. J	110	Lys	ALG		Set	vai	INI	ASI	GIY	rys	GIA	GIÀ	
		35					40			, .		45	:			
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cta	att	cat	crat	a++	det	++~	2+0					gtt				
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rea	TTE	Asp	Asp	IIe	Ala	Leu	Ile	Arg	Asp	Asp	Asp	Val	Leu	Phe	Val	
	50					55					60					
		•						. "			•					
			;			100	• • • •				4.7					
tgt	gaa	gga	gag	cca	ttt	att	gat	cct	cag	aca	gat	tct	aaq	cct	cct	421
Cys	Glu	Glv	Glu	Pro	Phe	Ile	Asp	Pro	Gln	Thr	Yen	Ser	Tara	Dro	Dwo	
65		- 4							· · · · ·		-ap	Ser	пys	PLO		
0.5					70					75					80	
gag	gga	tta	tta	gga	ttc	cac	aca	gac	taa	cta	202	tta		~++		460
G1.	Glv	Tau	T 011	23-	Dha	174 -	mb	3	-99	-		-	aac	guu	994	469
GIU	GIY	neu	Leu	GIY	Pne	HIS	Thr	Asp	Trp	Leu	Thr	Leu	Asn	Val	Gly	
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Gly	Arg	Tyr	Phe	Thr	Thr	Thr	Arg	Ser	Thr	Leu	Val	Asn	Lvs	Glu	Pro	
		-	100				_	105					110			
		٠.				2		203	•		1		110		•	
							•						-			
gac	agt	atg	ċtg	gċc	cac	atg	ttt	aag	gac	aaa	ggt	gtc	taa	qqa	aat	565
Asp	Ser	Met	Leu	Ala	His	Met	Dhe	Tare	Agn	Laze	611	Val	T	C1	200	
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aaσ	caa	gat	cár	aga	σσa	act	++6	tta	a++	<b>737</b>		agt		~		673
Tue	21-	3	***		22~	300		-		gac	cya	ayı	-	gag	Lac	613
Lys	GIII	ABD	HIS	Arg	GIY	ALA	Pne	Leu	Ile	Asp	Arg	Ser	Pro	Glu	Tyr	
	130					135	v*.				140					
		*	• •				٠.			٠,		•				
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CCC	gaa	CCC	att	ttg	aac	tac	ttg	cgt	cat	gga	cag	ctc	att	gta	aat	661
Phe	Glu	Pro	Ile	Leu	Asn	Tyr	Leu	Arg	His	Glv	Gln	Leu	Ile	Val	Asn	
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143		:			130		* "	1.		155					160	
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gat	ggc	att	aat	tta	tta	gat	ata	tta	gaa	gaa	ac=	aga	+++	+++	aat	709
A ===	Glv	T1 ~	7 ~-	T	7 644	01	373	*	520	27.0	3-0	. <u>~</u>			33.	,09
بإدم	GT Å	, <b>+</b> + =	Mail		hen	GTA	val	ren			Ala	Arg	hue	hue	GTÅ	
				165					170					175		
	. :			-								٠, .			•	
a + +	مدنى	+	4.34							· · ·	: ' '	· ·				
a	yac.	cca	cca	act	gaa	cac	cta	gaa	grg	gca	ata	aag	aat	țct.	caa	757
Ile	Asp	Ser	Leu	Ile	Glu	His	Leu	Glu	Val	Ala.	Ile	Lys	Asn	Ser	Gln	
			180		•			185		;			190			
1.7		. :	75.					200				٠.	.130			
7 2		٠.						u a i						3 ·	:	
cca	ccg	gag	gat	cat	tca	cca	ata	tcc	cga	aag	gaa	ttt	qtc	cqa	ttt	805

Pro Pro Glu Asp	His Ser Pro	Ile Ser Arg Lys 200	Glu Phe Val Arg Phe 205	
ttg cta gca act Leu Leu Ala Thr 210	cca acc aag Pro Thr Lys 215	tca gaa ctg cga Ser Glu Leu Arg	tgc cag ggt ttg aac Cys Gln Gly Leu Asn 220	853
ttc agt ggt gct Phe Ser Gly Ala 225	gat ctt tct Asp Leu Ser 230	cgt ttg gac ctt Arg Leu Asp Leu 235	cga tac att aac ttc Arg Tyr Ile Asn Phe 240	901
aaa atg gcc aat Lys Met Ala Asn	tta agc cgc Leu Ser Arg 245	tgt aat ctt gca Cys Asn Leu Ala 250	cat gca aat ctt tgc His Ala Asn Leu Cys 255	949
tgt gca aat ctt Cys Ala Asn Leu 260	gaa cga gct Glu Arg Ala	gat ctc tct gga Asp Leu Ser Gly 265	tca gtg ctt gac tgt Ser Val Leu Asp Cys 270	997
gcg aat ctc cag Ala Asn Leu Gln 275	Gly Val Lys	atg ctc tgt tct Met Leu Cys Ser 280	aat gca gaa gga gca Asn Ala Glu Gly Ala 285	1045
tcc ctg aaa ctg Ser Leu Lys Leu 290	tgt aat ttt Cys Asn Phe 295	gag gat cct tct Glu Asp Pro Ser	ggt ctt aaa gcc aat Gly Leu Lys Ala Asn 300	1093
tta gaa ggt gct Leu Glu Gly Ala 305	aat ctg aaa Asn Leu Lys 310	ggt gtg gat atg Gly Val Asp Met 315	gaa gga agt cag atg Glu Gly Ser Gln Met 320	1141 
aca gga att aac Thr Gly Ile Asn	ctg aga gtg Leu Arg Val 325	gct acc tta aaa Ala Thr Leu Lys 330	aat gca aag ttg aag Asn Ala Lys Leu Lys 335	1189
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aac gtg aag gga Asn Val Lys Gly 370	gct ata ttt Ala Ile Phe 375	gaa gag atg ctg Glu Glu Met Leu	aca cca cta cac atg Thr Pro Leu His Met 380	1333
Ser Gln Ser Val	aga t gagaat Arg		gaagatgtaa aagatgaaaa	1389
aaggaaattt taaaaaaaactga citti gtagggaaac tagai accaggcata gtati ggtttgagat gcati attgaattcc tagai aaggttgttc aggt	aaaaa cattta tittee atatte tattge tgeett ctatta tatttg ttgagg atttta tgeagt atggat ttataa ataget	gagg attatgettg tgat ttttaacaga ttga atggggtagg ettt taaataggca attt atggaaagca attt aaattgttaa ttag tgatgeetce	tctagaagaa ataacactgt ttttgagtgg tgcataaggg aaagcactca tttaatagat ggggtttacc tggtttatg tgatgtggaa ataccatctt caacatatgc aattatattt aactttatga aaacttggaa cctctttaaa tacctgtaac	1449 1509 1569 1629 1689 1749 1809 1869
gtttactttt tagg	acagaa cagtag	ctaa attaaagtaa	ttcaggttca tttttataat tatccagttc ttactgattg	1929

agacagagtg gaaagaaaga catcattgta catcactgtc attccaaagg tacagtgtaa ctctggatgg aggaataact tacctatcac tacaacactt acaaatgaga atttctcaga 2109 atttcattct aggcaagttc cactcaacac cagatcaagc aattctatct atttacacta 2169 ttagcctagt tttctcatac agtcatcaca agcataggaa gatacttcaa aaccaaaaaa accaaggtge atcattaata ttcatttaat tcaaatacca aatagtttac atagggccag cttagaaata gatactaaat ccagagctac tgcaatcaaa gcttatatga gtgaatatgg tagagttgcc tgctaaaagg caatgtaata taattgcagc tagaacccta cagtggggaa tgaggaattt taaacacaca tttgattaca gccaccaaaa aaatagacgt aaaaataaag gcatttggct ggtccaagat gtaattttca atcagtcagc acctgtgatt cttttactta tttttttgtg gtttttttt tttaaacaaa ttttagccca attttcttga gtcattctct ctctgcagca gcagaggaag ggcctgtacc tccctaccaa tgacttggtg tccttatttt ctaccccaag agcagggata ttagctgtgt ccaaatgggt tctgaattct acagactcat 2709 caacatgagg caaggaatca ttgaaaacca cctgtgtctc ctttgggaga atgacatatc 2769 tttagtattt acgtagetta ttettetata tetacatatg caaagettte ettaacagta 2829 aagggtacat atgcatagtg ggaggagatc agacctttac aagtgaagga aagcaacttc 2889 agaaatgaat tattttcttt gctttattat ttttaccaag acagagaagt attgtattga 2949 gagataatet atttteataa teaatatgtg eetaaattat atttaaatea ttteaetetg 3009 tactatattt tcaggaatta cagaatgtgg tattcattca cttaaaggta cctctgtaga 3069 aataacctaa aactgcagaa ggatctgaaa gatctaaaca tggtgtgctt agaaactgca 3129 gattttagat ctaatgtata ctgcattaat aaatgatata aagtgtttgt tgaaaaaaaa 3189 aaaaaaaaa aaaaa 3204

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THE DOOR STATE OF STATE

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260
                                                   270
Ala Asn Leu Gln Gly Val Lys Met Leu Cys Ser Asn Ala Glu Gly Ala
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Ser Leu Lys Leu Cys Asn Phe Glu Asp Pro Ser Gly Leu Lys Ala Asn
                       295
                                           300
Leu Glu Gly Ala Asn Leu Lys Gly Val Asp Met Glu Gly Ser Gln Met
305
                                       315
Thr Gly Ile Asn Leu Arg Val Ala Thr Leu Lys Asn Ala Lys Leu Lys
                325
                                   330
Asn Cys Asn Leu Arg Gly Ala Thr Leu Ala Gly Thr Asp Leu Glu Asn
         340
                               345
                                                   3.50
Cys Asp Leu Ser Gly Cys Asp Leu Gln Glu Ala Asn Leu Arg Gly Ser
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                                                                   120
ttctaatggc tycagetgey ctgggggety ggggctcccg ctgggactcc acttccgtgg
                                                                   180
atgtctaagc ttcacctttc ttgcgcccgc aggggcatga ctcaggtgaa agggagccat
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accgagecgt g atg etg ggg ttt gee atg atg gge tte tea gte eta atg
                                                                   470
            Met Leu Gly Phe Ala Met Met Gly Phe Ser Val Leu Met
ttc ttc ttg ctc gga aca acc att cta aag cct ttt atg ctc agc att
                                                                   518
Phe Phe Leu Leu Gly Thr Thr Ile Leu Lys Pro Phe Met Leu Ser Ile
                        20
cag aga gaa gaa teg ace tge act gee ate cae aca gat ate atg gae
                                                                   566
Gln Arg Glu Glu Ser Thr Cys Thr Ala Ile His Thr Asp Ile Met Asp
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                                        40
gac tgg ctg gac tgt gec ttc acc tgt ggt gtg cac tgc cac ggt cag
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Asp Trp Leu Asp Cys Ala Phe Thr Cys Gly Val His Cys His Gly Gln
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                                                       60
ggg aag tac ecg tgt ett eag gtg ttt gtg aac etc age eat eca ggt
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Gly Lys Tyr Pro Cys Leu Gln Val Phe Val Asn Leu Ser His Pro Gly
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                               70
cag aaa get ete eta eat tat aat gaa gag get gte eag ata aat eee
Gln Lys Ala Leu Leu His Tyr Asn Glu Glu Ala Val Gln Ile Asn Pro
                            85
                                 aag tgc ttt tac aca cct aag tgc cac caa gat aga aat gat ttg ctc
                                                                   758
                                  48
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Lys Cys Phe Tyr Thr Pro	Lys Cys His C	ln Asp Arg Asn i 105	Asp Leu Leu
aac agt gct ctg gac ata Asn Ser Ala Leu Asp Ile 110	: Lys Glu Phe F	tc gat cac aaa a he Asp His Lys i 120	aat gga act 806 Asn Gly Thr 125
ccc ttt tca tgc ttc tac Pro Phe Ser Cys Phe Tyr 130	Ser Pro Ala S	gc caa tct gaa g er Gln Ser Glu i 35	gat gtc att 854 Asp Val Ile 140
ctt ata aaa aag tat gad Leu Ile Lys Lys Tyr Asr 145	caa atg gct a Gln Met Ala I 150	le Phe His Cys	tta ttt tgg 902 Leu Phe Trp 155
cct tca ctg act ctg cta Pro Ser Leu Thr Leu Leu 160	ggt ggt gcc o Gly Gly Ala I 165	etg att gtt ggc a eu Ile Val Gly I 170	atg gtg aga 950 Met Val Arg
tta aca caa cac ctg tcc Leu Thr Gln His Leu Ser 175	tta ctg tgt g Leu Leu Cys 0	gaa aaa tat agc a Blu Lys Tyr Ser 1 185	act gta gtc 998 Thr Val Val
aga gat gag gta ggt gga Arg Asp Glu Val Gly Gly 190	' Lys Val Pro 1	at ata gaa cag o yr Ile Glu Gln 1 200	cat cag ttc 1046 His Gln Phe 205
aaa ctg tgc att atg agg Lys Leu Cys Ile Met Arg 210	Arg Ser Lys C	ga aga gca gag Bly Arg Ala Glu 115	aaa tct t 1092 Lys Ser 220
aagacggtgg ccaaattaaa g cctaattatg cctgtctgca a tcatgtggga aaaaaaaaa a	actaataat gtaa	aaggta ataattaa	tg caactgagga 1152 ag tatcatattt 1212 1246
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tgccaatgac agecttteet caccactaga tgtgcacaag	cctcaggga agaa	igagaga gacagact	ac agtgatggag 240

Met 1

347

 $\sqrt{(n_1)} \sum_{i=1}^n n_i^2 (q_i)$ 

ctg ggg ttt gcc atg atg ggc ttc tca gtc cta atg ttc ttc ttg ctc Leu Gly Phe Ala Met Met Gly Phe Ser Val Leu Met Phe Phe Leu Leu 5 10

gga aca acc att cta aag cct ttt atg ctc agc att cag aga gaa gaa 395 Gly Thr Thr Ile Leu Lys Pro Phe Met Leu Ser Ile Gln Arg Glu Glu 20 30

49

tcg	acc The	tgc	act	gcc	atc	cac	aca	gat	atc	atg	gac	gac	tgg	ctg	gac	443
Ser	Thr	cys	IIII	ALA	TIE	40	Thr	Asp	iie	Met	Asp 45	Asp	Trp	Leu	Asp	
	- :		•		•						_					
tgt	gcc	ttc	acc	tgt	ggt	gtg	cac	tgc	cac	ggt	cag	ggg	aag	tac	ccg	491
Cys 50	Ala	Pne	Thr	Cys	GLY 55	Val	His	Cys	His		Gln	Gly	Lys	Tyr		
50					23					. 60			• •		65	
tgt	ctt	cag	gtg	ttt	gtg	aac	ctc	agc	cat	cca	gat	cag	aaa	act	ctc	539
Cys	Leu	Gln	Val	Phe	Val	Asn	Leu	Ser	His	Pro	Gly	Gln	Lys	Ala	Leu	
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cta	cat	tat	aat	gaa	gag	act	atic	Cad	a+ a	aa⊭						505
Leu	His	Tyr	Asn	Glu	Glu	Ala	Val	Gln	Ile	Asn	Pro	Lvs	Cvs	Phe	Tyr	587
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						•				٠.					* 1	
Thr	cct	aag	tgc	Cac	Caa	gat	aga	aat	gat	ttg	ctc	aac	agt	gct	ctg	635
1111	Pro	100	Cys	nis	GIII	Asp	105	ASII	Asp	Leu	Leu	110	Ser	Ala	Leu	
• 1		D.				. •		:					• •			
gac	ata	aaa	gaa	ttc	ttc	gat	cac	aaa	aat	gga	act	ccc	ttt	tca	tgc	683
Asp	Ile	Lys	Glu	Phe	Phe	Asp	His	Lys	Asn	Gly	Thr	Pro	Phe	Ser	Cys	
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ttc	tac	ägt	cca	gcc	agc	caa	tċt	gaa	gat	ata	ått	ctt	ata		aac	731
Phe	Tyr	Ser	Pro	Ala	Ser	Gln	Ser	Glu	Asp	Val	Ile	Leu	Ile	Lys	Lys	/31
130					135					140				-	145	
+ a +	~~~	~~~	355	~~+									<i>:</i>			
Tyr	gac Asp	Gln	Met	Ala	Ile	Phe	His	Cvs	Len	Dhe	rgg	CCT	tca	ctg	act The	779
- 4 -				150				-75	155	FIIC	11p	PIO	261	160	1111	
		-							• ,							
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reu	Leu	GIA	165	Ala	Leu	TIE	vai	170	Met	Val	Arg	Leu		Gln	His	
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ctg	tcc	tta	ctg	tgt	gaa	aaa	tat	agc	act	gta	gtc	aga	gat	gag	gta	875
Leu	Ser-	Leu	Leu	Cys	Glu	Lys	Tyr	Ser	Thr	Val	Val	Àrg	Asp	Glu	Val	
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`aat	gga	aaa	qta	cct	tat	ata	gaa	cag	cat	cag	ttc	222	cta	tac	att	923
Gly	Gly	Lys	Val	Pro	Tyr	Ile	Glu	Gln	His.	Gln	Phe	Lvs	Leu	Cvs	Ile	723
	195					200					205	· -				
2+4	200	200	200		~~~						_	•				
Met	agg Arg	Ara	Ser	Lvs	Glv	Ara	Δla	Glu	Lve	Ser	t aa	ıgacg	gtgg	3		967
210							:			220						
					*			· · · ·								
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Asp	Cys 50		Phe	Thr	Cys	Gly 55		His	Cys	His	Gly 60		Gly	Lys	Tyr	
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Leu	Leu	His	Tyr	Asn 85	Glu	Glu	Ala	Val	Gln 90		Asn	Pro	Lys	Суs 95	Phe	
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		115			Phe		120					125	Pro			
	130				Ala	135					140				_	
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			180		Cys			185					1.90	_		
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TIE	210	Arg	Arg	ser	Lys	G1y 215	Arg	Ala	Glu	Lys	Ser 220	*	•			
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			DNA									٠.	•			
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                                              Met Arg Arg
ggc gcg ctt ctg gcg ggc gcc ttg gcc gcg tac gcg tac ctg gtg
                                                                  166
Gly Ala Leu Leu Ala Gly Ala Leu Ala Ala Tyr Ala Ala Tyr Leu Val
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ctg ggc gcg ctg ttg gtg gcg cgg ctg gag ggg ccg cac gaa gcc agg
                                                                  214
Leu Gly Ala Leu Leu Val Ala Arg Leu Glu Gly Pro His Glu Ala Arg
                                       30
etc ega gee gag etg gag aeg etg egg geg eag etg ett eag ege age
                                                                  262
Leu Arg Ala Glu Leu Glu Thr Leu Arg Ala Gln Leu Leu Gln Arg Ser
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ceg tgt gtg gct gcc ccc gcc ctg gac gcc ttc gtg gag cga gtg ctg
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Pro Cys Val Ala Ala Pro Ala Leu Asp Ala Phe Val Glu Arg Val Leu
                               60
gcg gcc gga cgg ctg ggg cgg gtc gtg ctt gct aac gct tcg ggg tcc
                                                                  358
Ala Ala Gly Arg Leu Gly Arg Val Val Leu Ala Asn Ala Ser Gly Ser
gcc aac gcc tcg gac ccc gcc tgg gac ttc gcc tct gct ctc ttc ttc
                                                                  406
Ala Asn Ala Ser Asp Pro Ala Trp Asp Phe Ala Ser Ala Leu Phe Phe
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gec age acg etg ate ace ace gtg gge tat ggg tae aca acg cea etg
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Ala Ser Thr Leu Ile Thr Thr Val Gly Tyr Gly Tyr Thr Thr Pro Leu
    105 116
act gat geg gge aag gee tte tee ate gee ttt geg ete etg gge gtg
                                                                  502
Thr Asp Ala Gly Lys Ala Phe Ser Ile Ala Phe Ala Leu Leu Gly Val
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ceg ace ace atg ctg ctg ace gec tea gec cag ege ctg tea etg
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Pro Thr Thr Met Leu Leu Leu Thr Ala Ser Ala Gln Arg Leu Ser Leu
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cgt Arg	ctc Lau	Ser 355	ttt Phe	Val	tcc Ser	cag Gln	cat His 360	ctg Leu	gct Ala	gly ggg	atg Met	tga *	agg Arg 365			1222
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235

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International application No. PCT/US99/03826

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	to International Patent Classification (IPC) or to both	national classification and IPC						
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Minimum de	ocumentation searched (classification system followe	d by classification symbols)						
U.S. :	636/23.1, 24.3; 435/7.2, 69.1, 320.1; 530/350							
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched								
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)								
Please See Extra Sheet.								
C. DOC	CUMENTS CONSIDERED TO BE RELEVANT							
Category*	Citation of document, with indication, where ap	propriate, of the relevant passages	Relevant to claim No.					
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International application No. PCT/US99/03826

#### **B. FIELDS SEARCHED**

Electronic data bases consulted (Name of data base and where practicable terms used):

APS, MEDLINE, JAPIO, BIOSIS, SCISEARCH, WPIDS, GENEMBL, NGENSEQ 34, EST, A-GENESEQ 32, PIR 58, SWISS-PROT 35, SPTREMBL 16.
search terms: potassium channel, K+hnov

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be searched, the appropriate additional search fees must be paid.

Group I, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:2, the nucleic acid having the sequence of SEQ ID NO:1, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:2 and K+Hnov protein of SEQ ID NO:2.

Group II, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:4, the nucleic acid having the sequence of SEQ ID NO:3, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:4 and K+Hnov protein of SEQ ID NO:4.

Group III, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:6, the nucleic acid having the sequence of SEQ ID NO:5, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:6 and K+Hnov protein of SEQ ID NO:6.

Group IV, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:8, the nucleic acid having the sequence of SEQ ID NO:7, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:8 and K+Hnov protein of SEQ ID NO:8.

Group V, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:10, the nucleic acid having the sequence of SEQ ID NO:9, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:10 and K+Hnov protein of SEQ ID NO:10.

Group VI, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:12, the nucleic acid having the sequence of SEQ ID NO:11, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:12 and K+Hnov protein of SEQ ID NO:12.

Group VII, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:14, the nucleic acid having the sequence of SEQ ID NO:13, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:14 and K+Hnov protein of SEQ ID NO:14.

Group VIII, claim(s)1-9, drawn to sucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:16, the sucleic acid having the sequence of SEQ ID NO:15, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:16 and K+Hnov protein of SEQ ID NO:16.

Group IX, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:18, the nucleic acid having the sequence of SEQ ID NO:17, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:18 and K+Hnov protein of SEQ ID NO:18.

Group X, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:20, the nucleic acid having the sequence of SEQ ID NO:19, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:20 and K+Hnov protein of SEQ ID NO:20.

Group XI, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:25, the nucleic acid having the sequence of SEQ ID NO:21-25, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:25 and K+Hnov protein of SEQ ID NO:25.

Group XII, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:27, the aucleic acid having the sequence of SEQ ID NO:26, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing

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K+Hnov protein of SEQ ID NO:27 and K+Hnov protein of SEQ ID NO:27.

Group XIII, claim(a)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:30, the nucleic acid having the sequence of SEQ ID NO:28-29, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:30 and K+Hnov protein of SEQ ID NO:30.

Group XIV, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:81, the nucleic acid having the sequence of SEQ ID NO:80, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:81 and K+Hnov protein of SEQ ID NO:81.

Group XV, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:83, the nucleic acid having the sequence of SEQ ID NO:82, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:83 and K+Hnov protein of SEQ ID NO:83.

Group XVI, claim(s)10, drawn to monoclonal antibody that binds to K+Hnov.

Group XVII, claim(s)11-14, drawn to non-human transgenic animal model for K+Hnov.

The inventions listed as Groups I-XVII do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: Group I is directed to nucleic acid (SEQ ID NO:1) encoding the K+Hnov protein of SEQ ID NO:2, nucleic acids hybridizing to said nucleic acid, expression cassette comprising said nucleic acid, cell comprising said cassette, method of producing the K+Hnov of SEQ ID NO:2 and the protein of SEQ ID NO:2. The special technical feature is the disclosed nucleic acid of SEQ ID NO:1 encoding the K+Hnov protein of SEQ ID NO:2. The nucleic acids, proteins, antibody and transgenic eximal model of Groups II-XVII do not share the special technical feature of Group I wherein the products of said Groups are structurally and functionally different. As shown in Table 1, pages 8-9, the H+Nov proteins of SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 25, 27, 30, 81 and 83 are all structurally and functionally different, the nucleic acids encoding said proteins having different chromosome positions.

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Box 1 Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)		
This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:		
1. [		Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
2. [		Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. [		Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box	11 (	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:		
	Ple	case See Extra Sheet.
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1. [		As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. [		As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. [		As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. X No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:  1-9, SEQ ID NO:1 and 2		
Rema	ırk o	The additional search fees were accompanied by the applicant's protest.
		No protest accompanied the new next of additional search fees.

Form PCT/ISA/210 (continuation of first sheet(1))(July 1992)\*